

Cell Surface Protein Biotinylation and Analysis

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[Abstract] A great way to specifically isolate and quantify proteins in the cell surface membrane is to take advantage of the biotinylation technique. It consists of labeling cell surface proteins with a biotin reagent before lysing the cells, and isolating these tagged proteins by NeutrAvidin pull-down. Then, the samples are subjected to SDS-PAGE separation, transferred to PVDF membranes and probed with specific antibodies. Quantification of cell surface expression is accomplished by densitometric measurement of the bands corresponding to the protein of interest and subsequent normalization by a membrane protein (as control).

Materials and Reagents

1. Human embryonic kidney HEK293 cells (Health Protection Agency Culture Collections, catalog number: 96121229)
2. Biotin reagent: EZ-link Sulfo-NHS-LC-LC-biotin (Thermo-Pierce, catalog number: 21338)
3. Electrophoresis grade Glycine (Sigma-Aldrich, catalog number: G8898)
4. Immobilized/NeutrAvidin Ultralink Resin (Thermo-Pierce, catalog number: 53150)
5. Pierce™ BCA Protein Assay Kit (Thermo-Pierce, catalog number: 23227)
6. Polyvinylidene difluoride (PVDF) membrane (GE Healthcare Life Sciences, catalog number: RPN303F)
7. Nonfat dry milk (AppliChem GmbH, catalog number: A0830)
8. Chemiluminescence substrate: Super signal west femto maximum sensitivity substrate (Thermo Fisher Scientific, catalog number: 34096)
9. X-ray films: Amersham Hyperfilm ECL (GE Healthcare Life Sciences, catalog number: 28-9068-35)
10. Primary antibodies

- a. Rabbit anti-human Nav1.5 antibody (Alomone Labs, catalog number: ASC-013)
- b. Mouse anti-Na⁺/K⁺ ATPase (Abcam, catalog number: ab7671)
11. Secondary antibodies
 - a. Stabilized Goat Anti-Mouse IgG (H + L), Peroxidase Conjugated (10 µg/ml) (Thermo Fisher Scientific, catalog number: 32430)
 - b. Stabilized Goat Anti-Rabbit IgG (H + L), Peroxidase Conjugated (10 µg/ml) (Thermo Fisher Scientific, catalog number: 32460)
12. Protease Inhibitors Cocktail (Roche Tablets, catalog number: 11836170001)
13. DPBS with calcium and magnesium (DPBS⁺) (see Recipes)
14. Lysis buffer LB1, LB2, LB3 (see Recipes)
15. Phosphate-Buffered Saline (PBS) (see Recipes)
16. Saline washing solution (SWS) (see Recipes)
17. 3x Gel loading buffer (see Recipes)
18. SDS-PAGE gel (see Recipes)
19. 5x Running buffer (see Recipes)
20. 10x Western blot transfer buffer (see Recipes)

Equipment

1. 35 mm Dish (Thermo Fisher Scientific, catalog number: 153066)
2. 6-well cell culture plate (BD Biosciences, catalog number: 353046)
3. Cell scrapers (VWR International, catalog number: 734-2603)
4. 96-well Microtest Plate (Sarstedt, catalog number: 82.1581)
5. Orbit™ LS Low Speed Shaker (LABNET, catalog number: S-2030-LS)
6. Rotating wheel (Noria R NR50, Ovan, catalog number: 10000-00062)
7. Microcentrifuge R5415 (Eppendorf, catalog number: 022621425)
8. Digital heat block (VWR International, catalog number: 460-3267)
9. Microplate Reader Benchmark plus (Bio-Rad Laboratories, catalog number: 170-6936)
10. Mini-PROTEAN Tetra Cell (Bio-Rad Laboratories, catalog number: 165-8000)
11. Mini Trans-Blot Module (Bio-Rad Laboratories, catalog number: 170-3935)
12. PowerPac HC Power Supply (Bio-Rad Laboratories, catalog number: 164-5052)

Software

1. ImageJ software (available at <http://rsb.info.nih.gov/ij>) (National Institute of Health, NIH)

Procedure

A. Biotinylation

Note: Perform all incubations on ice.

1. Remove growth media from the cells that you want to study, cultured in 35 mm plates.
Note: We used Human Embryonic Kidney HEK293 cells as experimental model. Cells were maintained in Dulbecco's Modified Eagle's Medium supplemented with 10% fetal bovine serum, 1% Penicillin-streptomycin and 1% Glutamax at 37 °C and 5% CO₂. We plated approximately 2.1×10^5 HEK293 cells in 35 mm dishes, but this value may vary depending on how fast your cell line grows (this will result in 70% of confluence 24 h later, which is the day of the transfection). Forty-eight hours after transfection we performed the Cell Surface Protein Biotinylation protocol.
2. Wash cells twice with 1 ml of ice cold DPBS⁺.
3. Incubate cells for 30 min on ice in the cold room with gentle rocking with 400 µl per plate of the biotin solution (2.5 mg/ml biotin reagent in DPBS⁺).
4. Wash each plate for 5 min three times with 1 ml of cold 100 mM Glycine in DPBS⁺, on ice in the cold room with gentle rocking.
5. Wash each plate for 5 min twice with 1 ml of cold 20 mM Glycine in DPBS⁺, as above.
6. Lyse with 200 µl of lysis buffer LB3 into each plate and use cell scrapers to detach the cells. Collect the cell lysates in 1.5 ml tubes.
7. For lysis, place the tubes in a rotating wheel at slow speed 1 h in the cold room.
8. While the lysis is performed, prepare the Immobilized/ NeutrAvidin Ultralink beads: For each sample, take 40 µl of NeutrAvidin beads at 50% slurry and wash the beads twice with 0.5 ml DPBS⁺ and twice again with 0.5 ml LB2 (for washing, centrifuge 30 sec at 3,000 x g at 4 °C and remove the supernatant). After the last wash, resuspend the precipitated beads with 20 µl of LB3.
9. Spin lysates at 16,000 x g (maximum speed) for 15 min at 4 °C.
10. Transfer the supernatants (solubilized material) to 1.5 ml tubes. Keep 10-15% of each supernatant in another tube and store at -80 °C (these are the INPUT samples, and the rest of the supernatant will be referred to as PULL-DOWN samples).
11. Incubate the pull-down samples with the 40 µl of NeutrAvidin beads prepared in step 8 overnight in the rotating wheel at slow speed in the cold room.
12. Centrifuge the samples at 16,000 x g, 30 sec at 4 °C.
13. Wash beads once with LB3, twice with LB2, twice with SWS and once with LB1. Each time, add 1 ml of the appropriate solution and centrifuge at 16,000 x g, 30 sec at 4 °C.
14. Resuspend the precipitated proteins in 25 µl of 2x gel loading buffer.
15. Heat the samples at 70 °C for 10 min in the heat block.

16. Keep pull-down samples at -20 °C until SDS-PAGE and Western blot.

B. Western blot

1. Use part of the input samples to quantify the protein using the BCA Protein Assay Reagent alongside BSA standards, following the manufacturer's directions. From each input sample, transfer equal amounts of protein to a new tube and mix with 3x gel loading buffer. Heat the samples at 70 °C for 10 min.

Note: Other protein quantification methods can be used and are equally efficient.

2. Load in an SDS-PAGE gel the aliquots of the inputs prepared in step B-1 and the total volume of the pull-down samples (without the beads). Before loading, spin down the samples to pellet the beads.
3. Run the gel at 40 V until samples have run through the stacking gel (30 min approximately). At that point, increase voltage to 100 V until the blue dye front has nearly run out of the gel.
4. Prepare the sandwich: Soak the gel 15 min shaking in Western blot transfer buffer. Activate the Polyvinylidene difluoride (PVDF) membrane by soaking it for 10 sec with methanol, then twice with water and 10 min with Western blot Transfer Buffer. Also, wet two sheets of filter paper and two fiber pads with Western blot Transfer Buffer. Construct the sandwich with the gel on the cathode side and the membrane on the anode side, with a sheet of filter paper and a fiber pad at each side.
5. Transfer the proteins to the PVDF membrane at 80 V for 2 h in the cold room.
6. Remove the membrane from the sandwich and block it with 5% nonfat milk in PBS with 0.1% Tween-20 1 h at room temperature.

Note: All Western blot incubations were performed in a shaker at low speed.

7. Incubate the membrane with primary antibody overnight at 4 °C.
Note: We used antibodies against human Na_v1.5 (protein of interest; rabbit polyclonal, at 1:1,000 in 5% non-fat milk in PBS with 0.1% Tween-20) and Na⁺/K⁺ ATPase (membrane protein used as control, mouse, at 1:5,000 in 5% non-fat milk in PBS with 0.1% Tween-20).
8. Wash three times with PBS with 0.1% Tween-20 to completely cover the membrane (approximately 20 ml), 10 min each, at room temperature.
9. Incubate with the appropriate secondary antibody 1 h at room temperature.
Note: We used peroxidase conjugated- goat anti-rabbit or anti-mouse IgG antibodies for Na_v1.5 and Na⁺/K⁺ ATPase, respectively (at 1:2,000 in 5% non-fat milk-PBS with 0.1% Tween-20).
10. Wash twice with PBS with 0.1% Tween-20 to completely cover the membrane (approximately 20 ml), 10 min each, at room temperature.

11. For signal development, use the chemiluminescence substrate and follow the manufacturer's instructions. Remove excess reagent and cover the membrane in transparent plastic wrap.
12. Detect the protein of interest by exposure to X-ray films (Figure 1).

Note: For normalization purposes, a membrane protein should also be assessed as control.

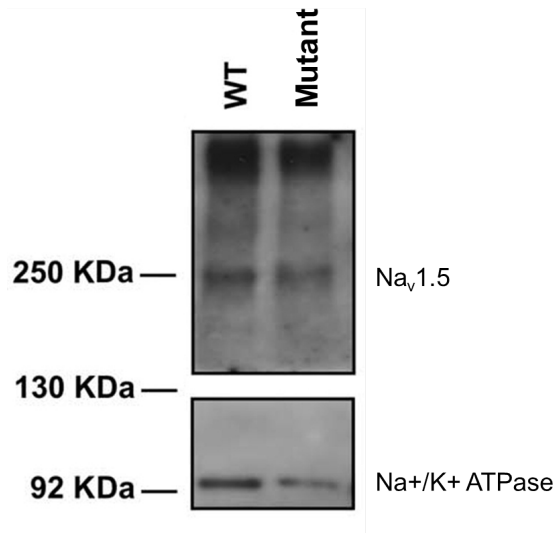


Figure 1. Western Blot detection of Na_v1.5 and Na⁺/K⁺ ATPase proteins performed after cell surface biotinylation from WT and mutant cells

C. Quantification

1. Scan the films in high resolution using the transparency mode.
2. Use the ImageJ software for band quantification. Follow the user's guide instructions to determine intensity values for each band as the integrated density (sum of pixel values) within a fixed area.

Note: When the aim is to compare WT and mutant conditions within multiple replicates, proceed as follows:

- a. Determine intensity values for the bands corresponding to WT and mutant conditions for the membrane protein used as control (IC_{WT} and IC_{MUT}, respectively) and WT and mutant condition for the protein of interest (I_{WT} and I_{Mut}, respectively).
- b. Calculate the ratio between IC_{WT} and IC_{MUT} to obtain the normalization factor (N_f):

$$IC_{WT}/IC_{MUT} = N_f$$

- c. Multiply I_{Mut} by the normalization factor calculated in the previous step to obtain the corrected mutant condition intensity value (I_{C Mut}):

$$I_{Mut} \times N_f = I_{C Mut}$$

- d. Divide $I_{C\text{ Mut}}$ by I_{WT} to obtain the expression of the mutant condition relative to I_{WT} .

$$I_{C\text{ Mut}}/I_{WT} = \text{mutant relative expression}$$

- e. Calculate the average of the mutant relative expression values for all the replicates and perform statistical analysis to assess possible differences with respect to the WT (Figure 2).

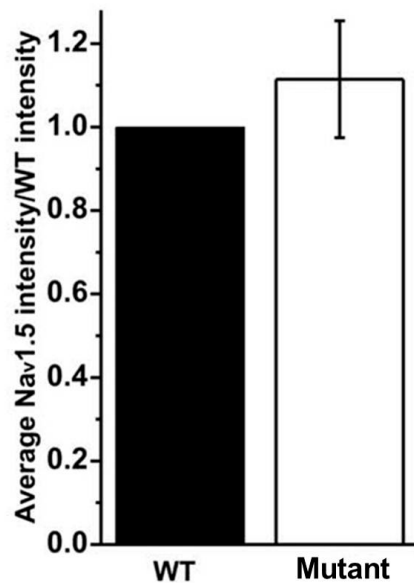


Figure 2. Bar graph shows the average of the intensity values obtained for each condition

Recipes

Note: All chemical reagents were obtained from Sigma-Aldrich unless stated otherwise.

1. DPBS⁺ (For 2 L)

0.2 g of CaCl_2

0.4 g KCl

0.4 g KH_2PO_4

0.2 g $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$

16 g NaCl

4.32 g $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$

To avoid forming an insoluble precipitate, add everything except CaCl_2 , and have it stirring in the cold room close to correct volume. While stirring and when cold, add CaCl_2 , and continue stirring for at least 10 min until the solution becomes clear. Add milli-Q water to the final volume of 2 L.

2. LB1
50 mM Tris/HCl (pH 7.4)
150 mM NaCl
1 mM EDTA
3. LB2
LB1 plus 1% (w/v) Triton X-100
To mix well, place the solution 1 h in cold room in the rotating wheel at slow speed.
4. LB3
For every 10 ml of solution LB2, add 1 tablet of Protease Inhibitors Cocktail right before use.
5. 1x PBS
8 g NaCl
0.2 g KCl
1.44 g Na₂HPO₄
0.24 g KH₂PO₄
Add milli-Q water for bring the final volume to 1 L.
6. SWS
0.1% Triton X-100 in PBS (pH 7.4)
350 mM NaCl
5 mM EDTA
7. 3x gel loading buffer
180 mM Tris/HCl (pH 6.8)
7.5% SDS
30% glycerol
0.051% Bromophenol blue
150 mM DTT
8. SDS-PAGE gel (for 2 gels 1.5 mm thick)
 - a. Stacking (4%)
4.65 ml H₂O
1.88 ml 0.5 M Tris (pH 7.4)
0.75 ml Acrylamide: Bisacrylamide 30% solution 37.5:1
75 µl 10% SDS
45 µl 10% Ammonium persulfate
15 µl N, N, N', N'-tetramethylethylenediamine (TEMED)
 - b. Resolving (4%)
9.35 ml H₂O
3.75 ml 1.5 M Tris (pH 8.8)

- 1.46 ml Acrylamide: 30% Bisacrylamide solution 37.5:1
- 150 µl 10% SDS
- 69 µl Ammonium persulfate 10%
- 23.1 µl N, N, N', N'-tetramethylethylenediamine (TEMED)
- 9. 5x Running buffer (For 1 L)
 - 5 g SDS
 - 144 g glycine
 - 30 g Tris
- 10. 10x Western blot transfer buffer (For 1 L)
 - 10 g SDS
 - 24.24 g Tris
 - 111.75 g glycine
 Before use, make up 1x and add 20% Methanol.

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