

# E15.5 Mouse Embryo Micro-CT Using a Bruker Skyscan 1172 Micro-CT

Elena Astanina<sup>1,2,\*</sup>, Sara Petrillo<sup>3</sup>, Tullio Genova<sup>4</sup>, Federico Mussano<sup>5</sup>, and Federico Bussolino<sup>1,2</sup>

<sup>1</sup>Department of Oncology, University of Torino, Torino, Italy

<sup>2</sup>Candiolo Cancer Institute-IRCCS-FPO, Candiolo, Italy

<sup>3</sup>Department of Molecular Biotechnology and Health Sciences, Molecular Biotechnology Center, University of Torino, Torino, Italy

<sup>4</sup>Department of Life Sciences and Systems Biology, University of Torino, Torino, Italy

<sup>5</sup>CIR Dental School, Department of Surgical Sciences, University of Torino, Via Nizza 230, 10126, Torino, Italy

\*For correspondence: [elena.astanina@ircc.it](mailto:elena.astanina@ircc.it)

## Abstract

X-ray computed microtomography ( $\mu$ CT) is a powerful tool to reveal the 3D structure of tissues and organs. Compared with the traditional sectioning, staining, and microscopy image acquisition, it allows a better understanding of the morphology and a precise morphometric analysis. Here, we describe a method for 3D visualization and morphometric analysis by  $\mu$ CT scanning of the embryonic heart of iodine-stained E15.5 mouse embryos.

**Keywords:** Imaging, X-ray computed tomography, Micro-CT, 3D visualization, Mouse embryo, Heart development

**This protocol was validated in:** Nat Commun (2022), DOI: 10.1038/s41467-022-32855-3

## Background

Transgenic mouse models have been extensively used to study the roles of specific genes in heart development. Historically, to analyze morphological defects on heart formation in gain-of-function and loss-of-function models, the formalin-fixed paraffin-embedded embryos were sectioned and analyzed section-by-section. X-ray computed microtomography ( $\mu$ CT) is a powerful tool for visualizing the 3D morphology of the heart and performing morphometric analysis.  $\mu$ CT uses X-rays to create cross-sections of a physical object that can be used to recreate 3D models, reaching very high resolution ( $<1\ \mu\text{m}$ ). Here, we describe a method to apply this technique to biological samples. Our purpose was to study the cardiovascular system in the developing mouse embryo. This method overcomes the limits of traditional histology, such as the difficulty to obtain comparable sections, especially when working with very small and complex samples.

For  $\mu$ CT analysis of embryos, it is mandatory to increase the contrast of tissues. To this end, different methods and contrast agents are described in the literature, with small differences in protocols. The main agents available to this purpose and tested by the authors are iodine solution or phosphotungstic acid (PTA) (Wong et al., 2012 and 2013; Hsu et al., 2016; Handschuh and Glösmann, 2022). Osmium tetroxide is also used as a contrast agent, but because of its high toxicity it has not been tested by the authors. Both approaches of staining were previously used to study cardiac defects (Degenhardt et al., 2010; Lesciotto et al., 2020; Jamet et al., 2022). In our experience, the use of 2%–5% PTA solution requires methanol (3%–10%) to properly penetrate tissues. The concentration of PTA and methanol can be adjusted depending on the size of sample; for example, for small mouse embryos (E12.5), 3% of methanol should be sufficient; for E15.5 embryos, 5% methanol; and for large mouse embryos (E18.5), 10% of methanol should be necessary. We successfully used this method for the staining of E13.5 mouse embryos (Petrillo et al., 2018). Although the PTA method offers excellent contrast, it may cause artifacts due to sample shrinkage in embryos older than E13.5. The maximum size of the object suitable for the  $\mu$ CT is 50 mm; however, increasing sample size will reduce the resolution.

On the contrary, iodine-based staining offers good balance in terms of sample preservation and soft tissue contrast, especially for larger samples. In our experiment, in E15.5 embryos we used a 20% Lugol's solution and 0.5% Tween-20. To preserve mineral structures, pH can be adjusted to 7.3.

## Materials and Reagents

### A. Mouse mating and embryo dissection

1. Petri dishes (100 mm) (BD Biosciences, catalog number: 351029 or similar)
2. 35 mm cell culture dishes (BD Biosciences, catalog number: 352096 or similar)
3. Sterile disposable tubes (50 mL) (BD Biosciences, catalog number: 352070 or similar)
4. 15 mL tubes
5. Mice
6. Pure ethanol (Sigma, catalog number: 493546)
7. Phosphate-buffered saline (PBS), without  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  (Life Technologies, Gibco®, catalog number: 10010)
8. 4% paraformaldehyde (PFA) (Santa Cruz Biotechnology, catalog number: sc-281692)
9. 70% ethanol (see Recipes)

### B. Micro-CT sample preparation

1. Lugol's solution (Sigma, catalog number: 32922)
2. Tween-20 (Sigma, catalog number: P1379)
3. Staining solution with contrast agent (see Recipes)

## Equipment

1. Blunt forceps (FST Standard Pattern forceps or similar)
2. Fine forceps (Dumont #5 or similar) and scissors (FST, catalog number: 14060-09 or similar)
3. X-Ray computed microtomography Skyscan 1172 (Bruker)

This instrument is a desktop  $\mu$ CT equipped with a fully distortion-corrected 11 MP X-ray camera (a 12-bit cooled CCD camera coupled to scintillator). The X-ray source is 20–100 kV, 10 W, with a  $<5\ \mu\text{m}$  spot size.  $0.8\ \mu\text{m}$  is the highest resolution (only for small samples) and  $25\ \mu\text{m}$  is the lower resolution. The maximum object size is 50 mm. Skyscan 1172 is equipped with three filter positions (None, 0.5 mm Al, or Al+Cu).

## Software

1. DataViewer 1.5.4.6 (Bruker)
2. NRecon 1.4.4 (Bruker)
3. CTVox 3.3.0 (Bruker)

## Procedure

### A. Mice mating

1. Keep mice on a 12:12 h light/dark cycle with unrestricted access to food and water. Use animals aged 2–4 months for mating.
2. For mating, put a male and two females in a cage after 6 pm.
3. Before 9 am on the next day, check for the formation of vaginal plug: lift a female by her tail, slightly dilate the vaginal opening with a blunt forceps, and check for a white mass.
4. Consider the day of plug detection as day E0.5. House the mated females separately from the male. Repeat mating procedure with non-mated females the following days.

### B. Embryo dissection

Embryo dissection is performed as described in Zeeb et al. (2012).

1. At day E15.5, euthanize the pregnant female.
2. Place the mouse on a supine position and spray 70% ethanol on the abdomen.
3. Make a V-formed incision on the skin and abdominal cavity and lift the tissue to expose the internal organs.
4. Identify the V-shaped uterus with the string of embryos inside, cut at the proximal (cervical) and distal (ovaries) ends, and remove the uterus to a tube with ice-cold PBS.
5. Dissect the embryos in a dish with PBS on ice. First, cut the uterus into single embryos-containing pieces. Place each embryo in a 35 mm dish with PBS and remove the uterine tissues and yolk sac. Keep yolk sac membranes for genotyping if required.

### C. Embryo fixation

1. Place single embryos in 15 mL tubes with 5 mL of 4% PFA and incubate at  $4\ ^\circ\text{C}$  for 24 h.
2. After fixation, keep the embryos in PBS at  $4\ ^\circ\text{C}$ . If needed, the samples can be kept at  $4\ ^\circ\text{C}$  for several days.

#### D. Micro-CT sample preparation

1. Prepare fresh staining solution of 20% Lugol's solution + 0.5% Tween-20 pH 7.3.
2. Keep samples in staining solution in constant gentle agitation.
3. Change solution daily for 10 days.

#### E. Micro-CT acquisition

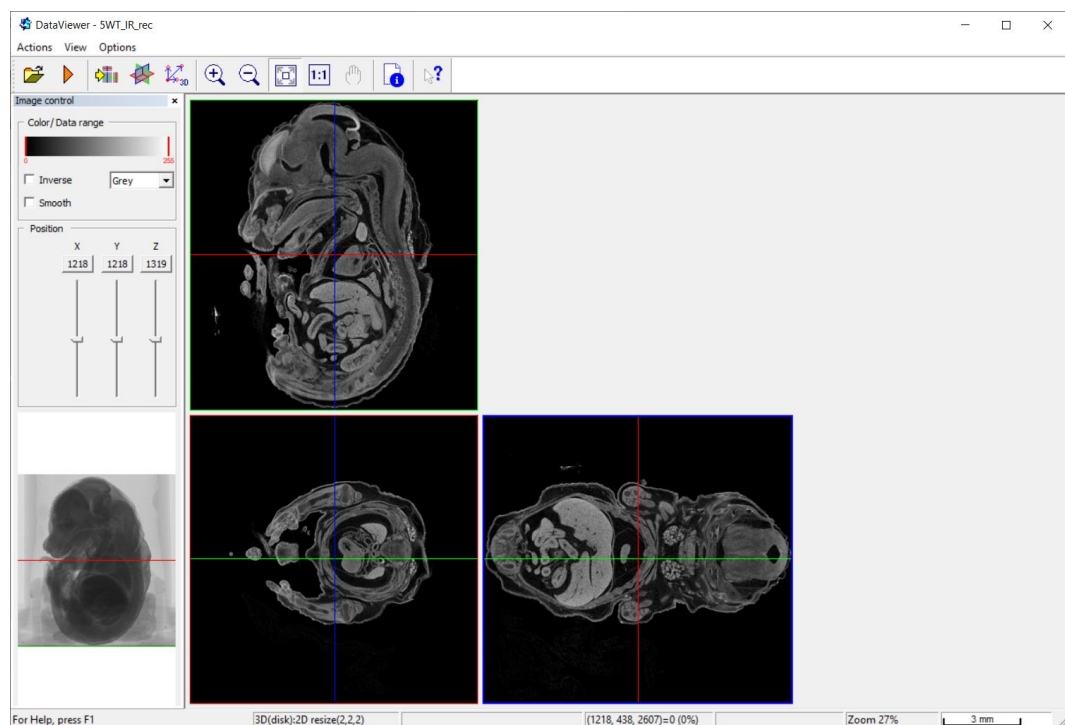
Perform acquisition at 80 kV using a 0.5 mm Al filter at a resolution of 3  $\mu\text{m}$ , 0.4° of rotation step, 360° scan, and 4× frame averaging.

#### F. Micro-CT image reconstruction

Obtain image reconstruction using NRecon software (Bruker), with proper misalignment compensation, no smoothing, 5 ring artifacts reduction, and 30% beam-hardening correction.

### Data analysis

1. Open dataset of the single embryo in DataViewer software (Figure 1).



**Figure 1. A screenshot of DataViewer software.** An embryo is visualized in virtual sections in three planes.

2. Select the plane by moving along the x- and y-axes. In our case, we were interested in performing the morphometric analysis of the embryo heart, in particular to measure the width of the ventricular walls and interventricular septum and the area of the pericardial cavity. The same plane, the best at representing the features we were interested in, was selected in all analyzed embryos.
3. Measure the features of interest by right-clicking to obtain the length in  $\mu\text{m}$ .

4. To perform a qualitative investigation, the dataset could be visualized in 3D by using CTVox software (Figure 2 and Video 1).

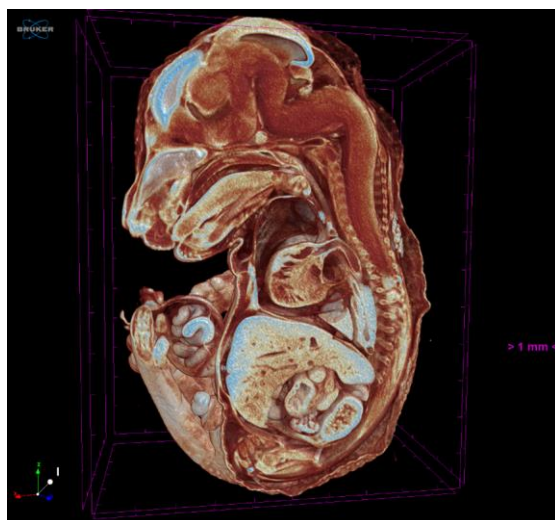


Figure 2. A screenshot of CTVox software. A E15.5 embryo is visualized in 3D slicing.



Video 1. Embryo

## Recipes

### 1. 70% ethanol

Obtained by mixing 70 mL of pure ethanol with 30 mL of water.

### 2. Staining solution with contrast agent

Staining solution with contrast agent is obtained by making a 20% Lugol's solution with 0.5% Tween-20 and adjusting pH to 7.3.

## Acknowledgments

This protocol was adapted from the previously published study (Astanina et al., 2022). This work was supported by AIRC – Associazione Italiana Per la Ricerca sul Cancro (grants 22910), Regione Piemonte (grant A1907A, Deflect), Fondazione CRT, Ministero dell'Università e della Ricerca (PRIN 2017, grant 2017237P5X), FPRC 5xmille 2016 MIUR (Biofilm), and ERA-Net Transcan-2 (grant TRS-2018-00000689) to F.B. and Ministero dell'Università.

## Competing interests

The authors declare no conflict of interests.

## Ethics

All animal procedures were approved by the ethics committee of the University of Turin and by the Italian Ministry of Health (protocol approval no. 864/2015-PR).

## References

- Astanina, E., Doronzo, G., Corà D., Neri, F., Oliviero, S., Genova, T., Mussano, F., Middonti, E., Vallariello, E., Cencioni, C., et al. (2022). [The TFEB-TGIF1 axis regulates EMT in mouse epicardial cells](#). *Nat Commun* 13(1): 5191.
- Degenhardt, K., Wright, A. C., Horng, D., Padmanabhan, A. and Epstein, J. A. (2010). [Rapid 3D phenotyping of cardiovascular development in mouse embryos by micro-CT with iodine staining](#). *Circ Cardiovasc Imaging* 3(3): 314-322.
- Handschuh, S. and Glösmann, M. (2022). [Mouse embryo phenotyping using X-ray microCT](#). *Front Cell Dev Biol* 10: 949184.
- Hsu, C. W., Wong, L., Rasmussen, T. L., Kalaga, S., McElwee, M. L., Keith, L. C., Bohat, R., Seavitt, J. R., Beaudet, A. L. and Dickinson, M. E. (2016). [Three-dimensional microCT imaging of mouse development from early post-implantation to early postnatal stages](#). *Dev Biol* 419(2): 229-236.
- Jamet, S., Ha, S., Ho, T. H., Houghtaling, S., Timms, A., Yu, K., Paquette, A., Maga, A. M., Greene, N. D. E. and Beier, D. R. (2022). [The arginine methyltransferase \*Carm1\* is necessary for heart development](#). *G3 (Bethesda)* 12(8).
- Lesciotta, K. M., Motch Perrine, S. M., Kawasaki, M., Stecko, T., Ryan, T. M., Kawasaki, K. and Richtsmeier, J. T. (2020). [Phosphotungstic acid-enhanced microCT: Optimized protocols for embryonic and early postnatal mice](#). *Dev Dyn* 249(4): 573-585.
- Petrillo, S., Chiabrando, D., Genova, T., Fiorito, V., Ingoglia, G., Vinchi, F., Mussano, F., Carossa, S., Silengo, L., Altruda, F., et al. (2018). [Heme accumulation in endothelial cells impairs angiogenesis by triggering paraptosis](#). *Cell Death Differ* 25(3): 573-588.
- Wong, M. D., Dorr, A. E., Walls, J. R., Lerch, J. P. and Henkelman, R. M. (2012). [A novel 3D mouse embryo atlas based on micro-CT](#). *Development* 139(17): 3248-3256.
- Wong, M. D., Spring, S. and Henkelman, R. M. (2013). [Structural stabilization of tissue for embryo phenotyping using micro-CT with iodine staining](#). *PLoS One* 8(12): e84321.
- Zeeb, M., Axnick, J., Planas-Paz, L., Hartmann, T., Strilic, B. and Lammert, E. (2012). [Pharmacological manipulation of blood and lymphatic vascularization in ex vivo-cultured mouse embryos](#). *Nat Protoc* 7(11): 1970-1982.