

# Measurement of Secreted Embryonic Alkaline Phosphatase

Meiyan Wang, Xinyi Wang, and Haifeng Ye\*

Synthetic Biology and Biomedical Engineering Laboratory, Biomedical Synthetic Biology Research Center, Shanghai Key Laboratory of Regulatory Biology, Institute of Biomedical Sciences and School of Life Sciences, East China Normal University, Dongchuan Road 500, Shanghai 200241, China

\*For correspondence: [hfy@bio.ecnu.edu.cn](mailto:hfy@bio.ecnu.edu.cn)

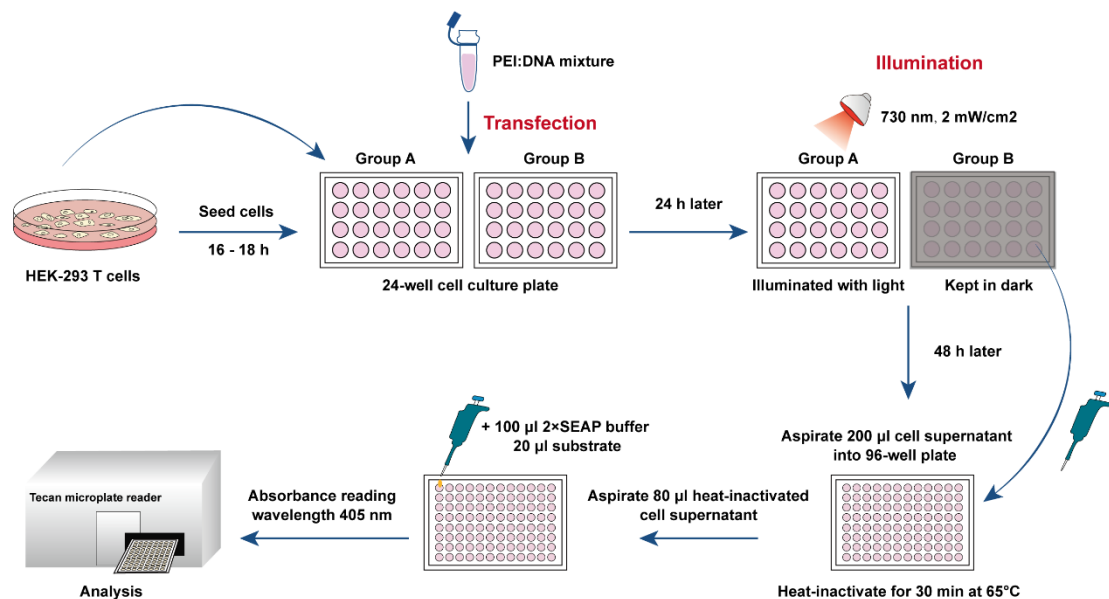
## Abstract

Secreted reporters have been demonstrated to be simple and useful tools for analyzing transcriptional regulation in mammalian cells. The distinctive feature of these assays is the ability to detect reporter gene expression in the culture supernatant without affecting the cell physiology or leading to cell lysis, which allows repeated experimentation and sampling of the culture medium using the same cell cultures. Secreted embryonic alkaline phosphatase (SEAP) is one of the most widely used reporter, which can be easily detected using colorimetry following incubation with a substrate, such as *p*-nitrophenol phosphate. In this report, we present detailed procedures for detection and quantification of the SEAP reporter. We believe that this step-by-step protocol can be easily used by researchers to monitor and measure molecular genetic events in a variety of mammalian cells due to its simplicity and ease of handling.

**Keywords:** SEAP, Secreted reporter, Reporter gene assays, Gene expression, Gene regulation

**This protocol was validated in:** Sci Adv (2021), DOI: 10.1126/sciadv.abh2358

## Graphical abstract



Schematic overview of the workflow described in this protocol

## Background

Reporter gene assays have been important for biomedical and pharmaceutical studies to monitor cellular activities associated with gene expression, regulation, and signal transduction. An ideal secreted reporter system would provide a more simple, rapid, and sensitive method to detect gene expression in a quantitative manner compared to classical intracellular reporters. Secreted embryonic alkaline phosphatase (SEAP), a C-terminal truncated variant of human placental alkaline phosphatase (PLAP), is one of the most widely used secreted mammalian reporter enzymes, engineered by deleting the 30 amino acid anchoring domain of PLAP (Berger et al., 1988). The SEAP reporter assay has superior properties: it is a simple and non-invasive procedure that does not require cell lysis and allows for long-term monitoring of gene expression in vitro and in vivo (Yang et al., 1997; Schlatter et al., 2002; Jiang et al., 2008). Moreover, transfected cells are not disturbed and remain intact for further studies during measurement of SEAP activity (Hu et al., 2022). Furthermore, the heat stability of SEAP and resistance to the phosphatase inhibitor L-homoarginine allow the samples to be pretreated at 65°C or with this inhibitor, to eliminate endogenous alkaline phosphatase activity (Tannous and Teng, 2011). Here, we describe the experimental procedures of in vitro measurement of SEAP adapted from our previously published work (Wang et al., 2021). This method can be useful for real-time monitoring of gene expression patterns in various cells and animals.

## Materials and Reagents

1. 96-well plate, transparent (Corning, catalog number: 3365)
2. 24-well plate, transparent (Corning, catalog number: 3537)
3. T25 flask (Corning, catalog number: 430639)
4. HEK-293T (ATCC, CRL-11268)
5. Dulbecco's modified Eagle's medium (DMEM) (Gibco, catalog number: 31600-083)

6. Fetal bovine serum (FBS) (Gibco, catalog number: 10270-106)
7. 0.25% trypsin-EDTA
8. 1% (vol/vol) penicillin and streptomycin solution (Beyotime Inc., catalog number: ST488-1/ST488-2)
9. p-nitrophenylphosphate (Sangon Biotech, CAS: 333338-18-4)
10. L-homoarginine (Sangon Biotech, CAS: 1483-01-8)
11.  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$  (Sangon Biotech, CAS: 7791-18-6)
12. Diethanolamine (Sangon Biotech, CAS: 111-42-2)
13. NaCl (Sangon Biotech, CAS: 7647-14-5)
14. KCl (Sangon Biotech, CAS: 7647-40-7)
15.  $\text{Na}_2\text{HPO}_4$  (Sangon Biotech, CAS: 7558-79-4)
16.  $\text{KH}_2\text{PO}_4$  (Sangon Biotech, CAS: 7778-77-0)
17. HCl (Sinopharm Chemical Reagent Co., Ltd., CAS: 7647-01-0)
18. Polyethyleneimine (PEI, molecular weight 40,000, stock solution 1 mg/mL in ddH<sub>2</sub>O) (Polysciences, catalog number: 24765)
19. 1× PBS (see Recipes)
20. 2× SEAP assay buffer (see Recipes)
21. Substrate solution (see Recipes)
22. Plasmids (see Table 1)

**Table 1. Plasmids designed and used in this study**

Plasmid	Description and cloning strategy	Reference
pXY137	Constitutive mammalian BldD and BphS expression vector (ITR- $P_{\text{hCMV}}$ -p65-VP64-BldD-pA- $P_{\text{hCMV}}$ -BphS-P2A-YhjH-pA-ITR)	Shao et al. (2017)
pZQ5	Far-red light-inducible expression vector (2*crRNA binding site- $P_{\text{hCMVmin}}$ -SEAP-pA)	This work
pZQ28	Constitutive crRNA <sub>SEAP</sub> expression vector ( $P_{\text{U6}}$ -crRNA <sub>SEAP</sub> -pA)	This work
pZQ113	Constitutive mammalian $P_{\text{hCMV}}$ -driven expression vector ( $P_{\text{hCMV}}$ -scFv-45bp linker-p65-HSF1-pA)	This work
pZQ116	Far-red light-inducible expression vector ( $P_{\text{FRL2}}$ -dAsCas12a-NLS-10×GCN4-pA; $P_{\text{FRL3}}$ , (OwhiG) <sub>2</sub> - $P_{\text{hCMVmin}}$ )	This work

## Equipment

1. Water bath
2. CO<sub>2</sub> incubator
3. Constant temperature incubator (65 °C)
4. Synergy H1 hybrid multi-mode microplate reader (BioTek Instruments, Inc.)
5. Multipass pipette
6. Reagent reservoirs
7. Hemocytometer (Merck, catalog number: Z359629)

## Software

1. Excel (Microsoft)
2. Gen5 software (version: 2.04)

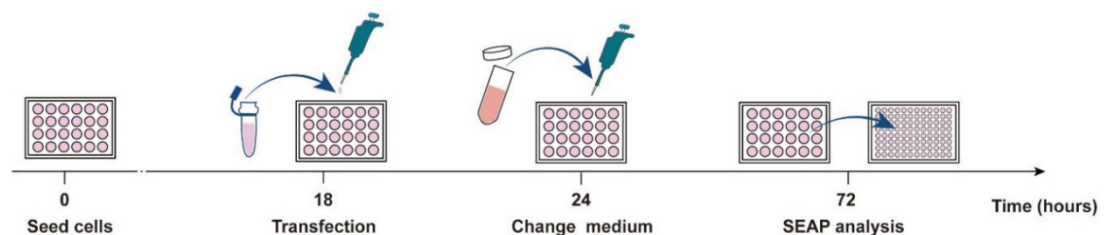
## Procedure

### Cell culture

1. Warm DMEM supplemented with 10% (vol/vol) FBS and 1% (vol/vol) penicillin and streptomycin solution (hereafter referred to as complete DMEM) and  $1\times$  PBS in a  $37^{\circ}\text{C}$  water bath for at least 15 min.
2. Wash ~90% confluent human embryonic kidney cells (HEK-293T) in a T25 flask with 3 mL of  $1\times$  PBS, aspirate the PBS, and add 0.5 mL of 0.25% trypsin-EDTA.
3. Incubate the cells at  $37^{\circ}\text{C}$  in a 5%  $\text{CO}_2$  incubator for 3–5 min, add 3 mL of prewarmed complete DMEM, and transfer the cell suspensions to a 15 mL sterile conical tube.
4. Spin down the cells at  $300\times g$  for 3 min at room temperature, discard the supernatant, and resuspend the cell pellet in 15 mL of complete DMEM.
5. Dispense 4 mL of the cell suspension into a T25 flask and incubate it for three days at  $37^{\circ}\text{C}$  in a humidified atmosphere containing 5%  $\text{CO}_2$  before subculturing again.

### Cell transfection

1. The 293T cell suspension should be prepared (as described in the Cell culture procedure) the day before transfection (Day 1) (**Figure 1**).

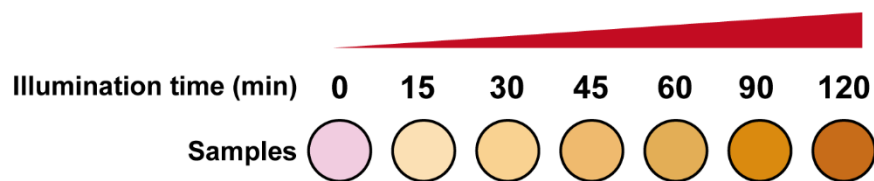


**Figure 1. Schematic diagram of the schedule for cell transfection and SEAP analysis**

2. Count the cells using a hemocytometer and seed  $6\times 10^4$  HEK-293T cells per well in a 24-well cell culture plate; culture for 18 h.
3. The next day (Day 2), prepare the transfection mix as follows:
  - a. Add 500 ng of the corresponding plasmid (see Table 1) into 50  $\mu\text{L}$  of FBS-free and antibiotic-free DMEM medium for each well of a 24-well plate.
  - b. Add 1.5  $\mu\text{L}$  of PEI (1  $\mu\text{g}/\mu\text{L}$ , PEI and DNA at a ratio of 3:1) into the above plasmid DNA solution.
  - c. To scale up, multiply the quantities by the number of replicates.
4. Vortex vigorously for 30 s.
5. Incubate the DNA-PEI mixture solution at room temperature for 15 min.
6. Add 50  $\mu\text{L}$  of transfection mix dropwise to each well. Rock the slide gently a few times and incubate at  $37^{\circ}\text{C}$  in a humidified atmosphere containing 5%  $\text{CO}_2$ .
7. Six hours after transfection, change the medium with fresh complete DMEM and incubate the 24-well plate at  $37^{\circ}\text{C}$  in a 5%  $\text{CO}_2$  incubator until Day 3.
8. On Day 3, the transfected slides can be used to perform SEAP reporter assay.

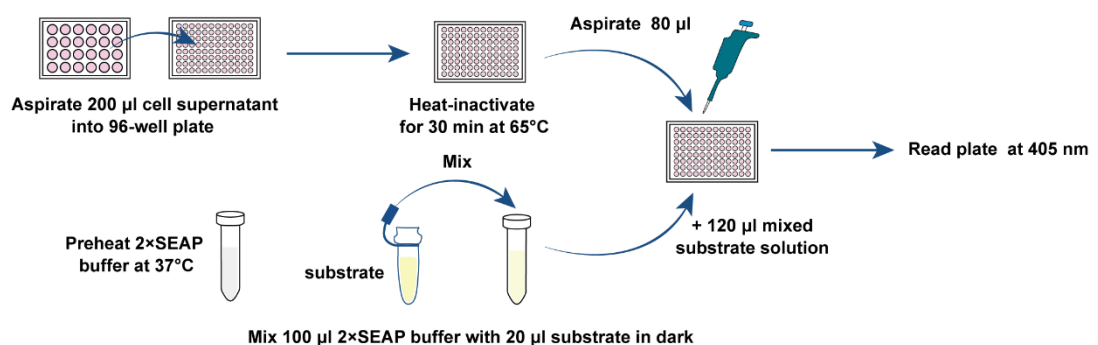
## SEAP reporter assay

1. Aspirate 200  $\mu$ L of cell culture supernatant from each sample and transfer into a chamber of a 96-well plate.
2. Heat-inactivate the cell culture supernatants at 65°C for 30 min.
3. Prewarm 2 $\times$  SEAP assay buffer (see Recipes) at 37°C and protect from light.
4. Mix 100  $\mu$ L of 2 $\times$  SEAP buffer with 20  $\mu$ L of substrate solution (see Recipes) in the dark.
5. Add 120  $\mu$ L of the above-mentioned mixed substrate solution into 80  $\mu$ L of heat-inactivated cell culture supernatant. At least three replicates should be performed for each group. The media will change color from pink to orange in the presence of SEAP (**Figure 2**).



**Figure 2. Detection of SEAP production at the indicated time points after illumination.** In the presence of SEAP, the media changes from pink to orange. The yellow color intensity correlates with light illumination time.

6. Measure light absorbance at 405 nm at 37°C for 15 min using a multi-mode microplate reader. See **Figure 3** for the detailed procedure.

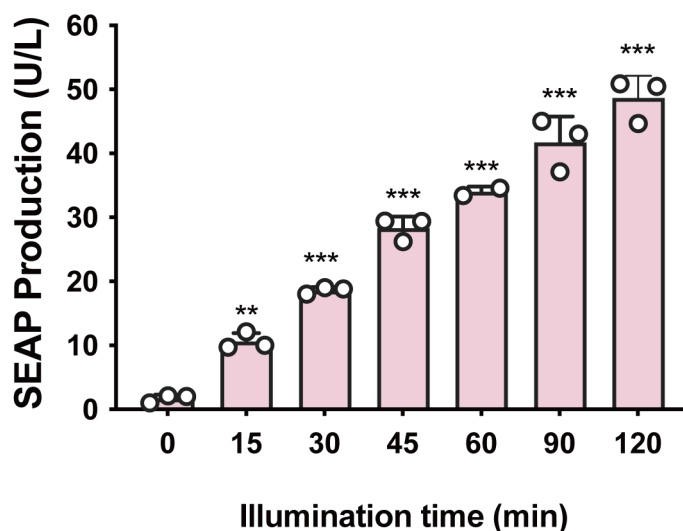


**Figure 3. Schematic diagram of SEAP reporter assay**

7. Calculate the levels of SEAP activity using data points that yield a rate of change in light absorbance that is linear with respect to time. (If available, a computer-linked kinetic ELISA plate reader is most convenient, as this will precisely calculate the linear change in  $A_{405}$  in all 96 wells of a microplate over time.)

## Data analysis

For each illumination time, data are expressed as mean  $\pm$  standard deviation (SD);  $n = 3$  independent experiments. The data can also be represented as fold change difference compared to non-stimulated cells and analyzed by one-way ANOVA followed by Tukey's post-hoc tests. Perform statistical analysis using GraphPad Prism or other software.



**Figure 4. Quantification of far-red light-inducible SEAP expression kinetics.** Transfected HEK-293T cells were illuminated with far-red light for different time periods (0–120 min). SEAP production in the culture supernatant was profiled at 24 h after illumination. \*\*P < 0.01, \*\*\*P < 0.001 illumination groups vs. 0 min.

## Notes

1. The mixture of 2× SEAP buffer and substrate solution should be protected from light, quickly added to cell culture supernatants, and detected immediately.
2. If SEAP production is too high, the samples should be diluted with PBS buffer solution.

## Recipes

### 1. 1× PBS buffer

Reagent	Final concentration	Amount
NaCl	137 mM	8.00 g
KCl	2.7 mM	0.20 g
Na <sub>2</sub> HPO <sub>4</sub>	10 mM	1.42 g
KH <sub>2</sub> PO <sub>4</sub>	1.8 mM	0.24 g
H <sub>2</sub> O	n/a	800 mL
Adjust pH to 7.4 with HCl, then add ddH <sub>2</sub> O to final volume (1,000 mL)		
Total	n/a	1,000 mL

## 2. 2× SEAP buffer

Reagent	Final concentration	Amount
L-homoarginine	20 mM	4.4938 g
MgCl <sub>2</sub> (3 M)	1 mM	0.334 mL
Diethanolamine	21% (v/v)	210 mL
Adjust pH to 9.8 with HCl, then add dH <sub>2</sub> O to final volume (1,000 mL)		
Total	n/a	1,000 mL

Store at 4°C and protect from light.

## 3. Substrate solution

Reagent	Final concentration	Amount
p-nitrophenylphosphate	120 mM	2.226 g
2× SEAP buffer	n/a	50 mL
Total	n/a	50 mL

Aliquot, store at -20°C, and protect from light.

## Acknowledgments

This protocol was adapted from previous work (Wang et al., 2021. DOI: 10.1126/sciadv.abh2358). This work was financially supported by grants from the National Key R&D Program of China (No. 2019YFA0904500, No. 2019YFA0110802), the National Natural Science Foundation of China (NSFC: No. 32171414, No. 31971346, No. 31861143016, No. 31870861), the Nature Science Foundation of Chongqing, China (No. CSTB2022NSCQ-MSX0461), the Open Project Program of State Key Laboratory of Dairy Biotechnology (No. SKLDB2021-001), and the Instruments Sharing Platform of the School of Life Sciences, East China Normal University.

## Competing interests

All authors wish to declare no competing interests.

## References

- Berger, J., Hauber, J., Hauber, R., Geiger, R. and Cullen, B. R. (1988). [Secreted placental alkaline phosphatase: a powerful new quantitative indicator of gene expression in eukaryotic cells](#). *Gene* 66(1): 1-10.
- Hu, Z., Zhang, T., Jiang, S. and Yin, H. (2022). [Protocol for evaluation and validation of TLR8 antagonists in HEK-Blue cells via secreted embryonic alkaline phosphatase assay](#). *STAR Protoc* 3(1): 101061.
- Jiang, T., Xing, B. and Rao, J. (2008). [Recent developments of biological reporter technology for detecting gene expression](#). *Biotechnol Genet Eng Rev* 25: 41-75.
- Schlatter, S., Rimann, M., Kelm, J. and Fussenegger, M. (2002). [SAMY, a novel mammalian reporter gene derived from \*Bacillus stearothermophilus\* alpha-amylase](#). *Gene* 282(1-2): 19-31.
- Shao, J., Xue, S., Yu, G., Yu, Y., Yang, X., Bai, Y., Zhu, S., Yang, L., Yin, J., Wang, Y., Liao, S., Guo, S., Xie, M., Fussenegger, M. and Ye, H. (2017). [Smartphone-controlled optogenetically engineered cells enable semiautomatic glucose homeostasis in diabetic mice](#). *Sci Transl Med* 9(387): eaal2298.

- Tannous, B. A. and Teng, J. (2011). [Secreted blood reporters: insights and applications](#). *Biotechnol Adv* 29(6): 997-1003.
- Wang, X., Dong, K., Kong, D., Zhou, Y., Yin, J., Cai, F., Wang, M. and Ye, H. (2021). [A far-red light-inducible CRISPR-Cas12a platform for remote-controlled genome editing and gene activation](#). *Science advances* 7(50): eabh2358.
- Yang, T. T., Sinai, P., Kitts, P. A. and Kain, S. R. (1997). [Quantification of gene expression with a secreted alkaline phosphatase reporter system](#). *Biotechniques* 23(6): 1110-1114.