

## Measuring Cell Growth and Junction Development in Epithelial Cells Using Electric Cell-Substrate Impedance Sensing (ECIS)

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**[Abstract]** Electric Cell-substrate Impedance Sensing (ECIS) is an automated method that can be used to quantify processes such as cell attachment, growth, migration and barrier functions (*i.e.*, the properties of tight junctions). The method provides simultaneous information on cell number and tight junction function by detecting electric parameters of cells grown on electrodes. Samples are probed with small alternating current (AC) over a range of frequencies, and changes in capacitance and impedance are measured over time. Capacitance reflects the degree of electrode coverage by cells, that correlates with cell number, and can be used to assess cell proliferation or migration. Impedance values inform about barrier function. Obtaining real-time simultaneous information on these parameters is unique to this system and is of great value for addressing fundamental questions such as the role of tight junction proteins in cell growth and migration. This protocol describes the use of ECIS to follow cell growth and tight junction-dependent barrier generation in tubular epithelial cells. We used this method to explore how depleting claudin-2, a tight junction protein affects tubular cell growth and barrier function. During the process, cells are transfected with control or claudin-2-specific siRNA, and 24h later plated on electrodes. ECIS automatically collects information on cell growth and barrier as the monolayer develops. The data are initially analyzed using the ECIS software and exported into a graph software for further processing.

**Keywords:** Impedance sensing, Epithelial cells, Cell growth, Transepithelial resistance, Tight junction development

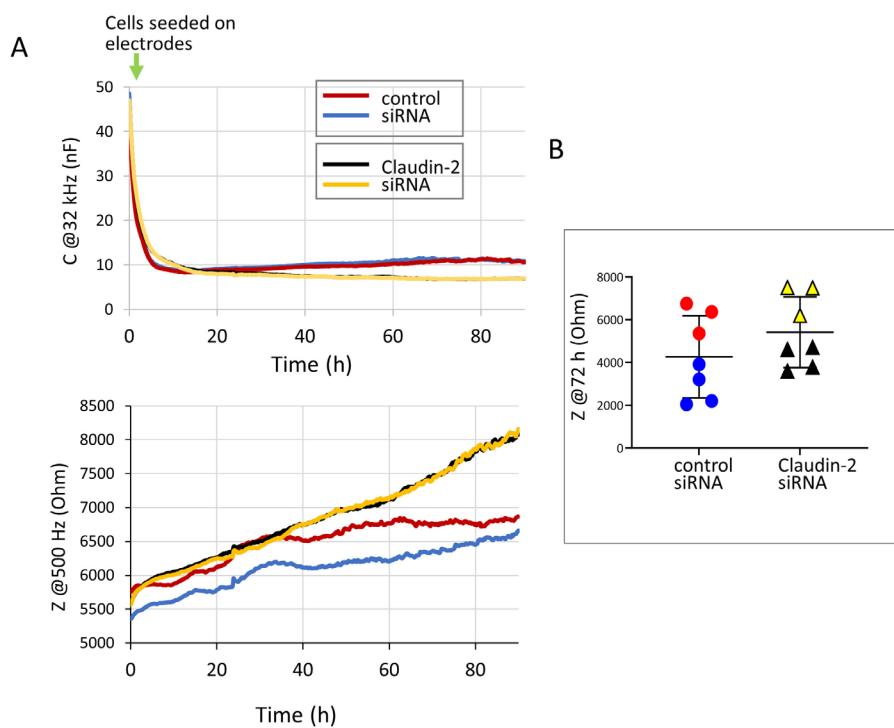
**[Background]** Epithelial cells generate monolayers characterized by well-developed intercellular junctions. Input from the junctions determines cell behavior, including proliferation and migration. As junctions develop, cell proliferation and migration slows and then halts due to contact inhibition (McClatchey and Yap, 2012). Although the past decades rapidly augmented our understanding of the molecular mechanisms of proliferation and migration, many aspects of these processes, and the exact nature of contact inhibition remain incompletely defined. As cells establish a confluent layer, intercellular junctions, including tight junctions develop and mature, ensuring well-controlled paracellular permeability (barrier functions) (Zihni *et al.*, 2016). Paracellular seal and permeability pathways are established by the claudin family of tight junction proteins (Tsukita *et al.*, 2019). Interestingly, accumulating evidence suggests that in addition to their role as permeability regulators, many claudins also modulate proliferation and migration (Hagen, 2017; Venugopal *et al.*, 2019). These roles, however,

remain incompletely understood. Methods that follow barrier function and cell growth or migration simultaneously over time are crucial for obtaining new insights. Electric Cell-substrate Impedance Sensing (ECIS), that was developed by Applied BioPhysics offers this possibility. ECIS is an impedance-based, automated method that allows the quantification of various aspects of cell behavior through detection of electric properties (Wegener *et al.*, 2000). The cells are grown in chambers containing small gold electrodes (referred to as arrays), and the system continuously probes the electrodes using small, biologically inert alternating current. A range of AC frequencies from 62.5-64,000 Hz is used to measure impedance and capacitance across the electrodes over time. The basis of the measurement is that the current can flow through the cells or under and between them, and the magnitude of resistance of these compartments depends on the frequency of the AC current applied. Thus, specific frequencies can be used to probe different parameters, and this can be utilized to follow various aspects of cell behavior in the same sample (Stolwijk *et al.*, 2015; Wegener *et al.*, 2000). During measurement the system measures capacitance and impedance at the full frequency range, and the user can select the required parameters during analysis. Capacitance detected using high AC frequency reflects coverage of the electrode, with little influence from other parameters. Capacitance data collected in a typical experiment are shown in Figure 1A (top graph). This demonstrates that following seeding cells generate an insulating layer on the electrodes that is reflected by a drop in capacitance. Capacitance reaches its minimum and stabilizes when a fully confluent layer is generated. The initial rate of drop reflects cell attachment, that is completed by 3h. After this time, the rate of drop reflects cell growth. Figure 1A bottom graph shows the parallel changes in impedance ( $Z$ ) values measured at low AC frequencies. In epithelial cells with well-developed tight junctions, impedance measured at 500 Hz or lower frequencies reflects predominantly the properties of the paracellular space. Impedance increases as cells reach confluence and tight junctions develop. Many epithelial cells show an increase in the  $Z$  value, followed by a decrease as junctions mature (Amoozadeh *et al.*, 2017). Interestingly, simultaneous measurement of capacitance and impedance reveals that tight junction development and maturation does not stop as confluence is reached (Amoozadeh *et al.*, 2018). More detailed analysis to separate different components of impedance can be performed using mathematical modelling, built into the software, which can be advantageous in cells with weaker paracellular resistance (e.g., endothelial cells) (Wegener *et al.*, 2000; Stolwijk *et al.*, 2015; García *et al.*, 2019). Although the theory of mathematical modelling in ECIS is complex, the measurement itself is relatively straight forward, and simple analysis can provide valuable information. ECIS is also equipped with an elevated field module for wounding of the layer, and capacitance values can be used for an automated cell migration assay (not discussed in this protocol) (Szaszi *et al.*, 2012).

The advantages of ECIS are as follows. It is an automated, continuous, real-time, label-free detection method. Since the arrays are housed in a tissue culture incubator, the environment is controlled and any measurement artifacts stemming from changes in temperature and  $\text{CO}_2$  levels are eliminated. In comparison with other methods that follow cell growth or migration, the key benefit of ECIS is the ability to simultaneously detect transepithelial resistance (TER), *i.e.*, barrier function and cell growth or migration (Kakiashvili *et al.*, 2011; Amoozadeh *et al.*, 2015 and 2017; Stolwijk *et al.*, 2015; Láng *et al.*,

2017) (Figure 1). This can be valuable for correlating effects of various treatments on these highly interdependent parameters, thereby providing new insights into fundamental cell biological questions.

This protocol describes the use of ECIS to follow tubular epithelial cell growth, and the development of barrier via formation of the tight junctions. We used this protocol to assess the development of cell confluence (Amoozadeh *et al.*, 2018) and to follow effects of silencing of a tight junction protein, claudin-2 rate on proliferation rate and barrier function (Dan *et al.*, 2019). Silencing claudin-2 alters both barrier function (resulting in elevated resistance) (Amoozadeh *et al.*, 2018) and cell growth (reducing proliferation) (Dan *et al.*, 2019). Simultaneous detection of these parameters reveals both effects. We found that claudin-2 silencing slows the development of confluence, but does not prevent it, and claudin-2 depleted cells generate a barrier which is tighter than the control due to absence of the channel forming claudin-2. Adaptation of the method for other cell types and proteins requires optimization of the seeding conditions, electrode coating and transfection parameters. We provide notes on these issues to direct the reader.



**Figure 1. Capacitance and impedance measurements of LLC-PK1 cells with or without claudin-2 silencing.** A. Exported graphs showing simultaneously detected changes in capacitance at 32 kHz (reflecting cell growth) and impedance (z) at 500 kHz (reflecting barrier). LLC-PK1 cells were transfected using control or claudin-2-specific siRNA and electrical properties were measured using ECIS, as described in the protocol. B. Impedance (z) was determined at 72 h after seeding of cells in two different experiments. The data from the same experiment are indicated by black and red for experiment #1 and blue and green for experiment #2 (control and claudin-2 siRNA, respectively).

## Materials and Reagents

1. Clear tissue culture-treated 6-well microplates (Corning Costar®, Millipore-Sigma, catalog number: CLS3513)
2. Microcentrifuge tubes (1.5 ml) (Fisherbrand, catalog number: 2043-05408129)
3. Countess™ Cell Counting Chamber Slides (Thermo Fisher, Invitrogen, catalog number: C10228)
4. Cell line: LLC-PK<sub>1</sub> kidney tubule epithelial cell line (European Collection of Authenticated Cell Cultures, catalog number: 86121112)
5. Cell culture media and reagents:
  - Dulbecco's Modified Eagle Medium (D-MEM), low glucose, with L-glutamine and 110 mg/L sodium pyruvate (Gibco, Thermo Fisher, catalog number: 11885084)
  - Fetal Bovine Serum (Gibco, Thermo Fisher, catalog number: 12483-020)
  - Penicillin-Streptomycin, 100x, sterile-filtered, cell culture tested (Sigma-Aldrich, catalog number: P-4333. 25300062)
  - Trypsin-EDTA (0.05% Trypsin with EDTA 4Na) (Thermo Fisher, Gibco, catalog number 25300062)
  - Opti-MEM™ I Reduced Serum Medium for transfection (Thermo Fisher, Gibco, catalog number: 31985088)
6. Trypan Blue Stain (0.4%) (Gibco, catalog number: 15250061) for use with the Countess™ Automated Cell Counter (Invitrogen, Thermo Fisher, catalog number T10282)
7. Lipofectamine™ RNAiMAX Transfection Reagent (Invitrogen, Thermo Fisher, catalog number 13778150)
8. Two custom-designed siRNAs targeting porcine claudin-2 (Thermo Fisher/Dharmacon) (sequence #1, CCAGAACTCTCGCGCCAAA, and #2 CCCTGATAGCTGGGATCAT)
9. Silencer™ Negative Control No. 1 siRNA (non-related (NR)) siRNA (Applied Biosystems/Ambion, Thermo Fisher, catalog number: AM4611)
10. ECIS electrode array, type: 8W10E or 8W10E+ PET (Applied Biophysics, catalog number: 8W10E or 8W10E+)
11. L-cysteine (Millipore-Sigma-Aldrich catalog number: C7352)
12. 10 mM L-cysteine solution for electrode stabilization (see Recipes)

## Equipment

1. ECIS® Z-Theta instrument with 16-well array module housed in incubator (Applied Biophysics)
2. Countess™ Automated Cell Counter (Thermo Fisher, Invitrogen, catalog number C10228)
3. Tissue culture equipment (sterile hood, incubator, centrifuge)

## Software

1. ECIS Zθ software
2. Graph software (e.g., Excel or GraphPad)

## Procedure

### A. Electrode stabilization with cysteine pretreatment

1. Pretreat electrodes with 200  $\mu$ l of filtered 10 mM L-cysteine for 15 min at room temperature.
2. Aspirate the cysteine solution and wash with 200  $\mu$ l of autoclaved dH<sub>2</sub>O twice. Do not wash electrodes with PBS.
3. Air dry the electrodes before seeding cells. Alternatively, the treated and dried electrodes can be stored at room temperature till its expiry.

### B. Splitting epithelial cells for transfection with siRNA

1. Grow LLC-PK<sub>1</sub> cells in low glucose DMEM supplemented with 10% FBS and 1% Pen-Strep in an incubator at 37 °C with 5% CO<sub>2</sub>.
2. Split cells when they are grown to about 90% confluence.
3. Seed cells for transfection in 6-well plates using standard cell culture procedures.
4. Use 2 ml antibiotic-free, serum-containing DMEM/well to seed cells. Aim at obtaining 30-40% confluence the next day.

### C. Transfecting cells

The following day (16-24 h later) transfect cells with 100 nM of the negative control (non-related, NR) or claudin-2-specific siRNA using the Lipofectamine RNAiMAX transfection reagent as follows:

#### 1. Mix reagents:

Tube A: For each condition (well) to be transfected, prepare siRNA: dilute 2.4  $\mu$ l of 100 nM stock siRNA duplex in 200  $\mu$ l Opti-MEM medium without serum in a sterile microcentrifuge tube. Mix gently. For more wells, scale up amounts as needed.

Tube B: For each condition (well) prepare Lipofectamine<sup>TM</sup>: add 3  $\mu$ l Lipofectamine<sup>TM</sup> RNAiMAX to 200  $\mu$ l Opti-MEM medium. Mix gently. For more wells, scale up amounts as needed.

2. Incubate the tubes for 5 min at room temperature.
3. Mix the content of tube A and B to generate the siRNA-Lipofectamine complex. Incubate for 15 min.
4. At the end of the incubation time, add 400  $\mu$ l of the siRNA-Lipofectamine mix to the corresponding well. Gently shake to mix and return to the incubator.
5. Grow cells for 24 h.

#### D. Seeding cells on the electrode arrays

1. The next day, prepare the electrode arrays for seeding:

Add 400  $\mu$ l pre-warmed complete culture medium (growth medium with serum and antibiotics) to the wells, and place into the incubator for 15 min prior to seeding cells. This will coat the surface with serum and warm the array to reduce convection of the medium/cell suspension due to heat differences, that affects even settling of cells on the electrode.

2. Trypsinize cells transfected with control siRNA and claudin-2-specific siRNA. When cells are floating, add warm medium, collect, and spin down cells using 300  $\times$  g. Resuspend in fresh medium. Keep warm while performing the cell counting.

3. Count cells using the Countess automated cell counter:

Using a sterile tip remove 10  $\mu$ l of the cell suspension and mix with 10  $\mu$ l of Trypan blue in a microcentrifuge tube. Add to a Countess<sup>TM</sup> Cell Counting Chamber Slide and count cells using the automated cell counter to obtain number of viable cells.

*Note: Manual counting can be used instead of the Countess.*

4. Adjust volume using warm medium to obtain a total of  $0.5 \times 10^5$  cells/electrode in a final volume of 400  $\mu$ l.

5. Remove array from the incubator and aspirate the medium from the wells. Add 400  $\mu$ l of cell suspension to each well.

6. Insert the electrode into the ECIS array holder in the incubator and screw the holder tight.

#### E. Software setup and acquiring data

1. Turn on the computer and the ECIS Z0 machine.

2. Open the ECIS software. Click the “Setup” button. The system will check the connection. If the connection is good, the wells show up as green. If the connection is bad (red wells), reinsert the electrode, then click “Check” to confirm connectivity of all electrodes by using the “Check” button.

3. Select the type of electrode. Choose the “Multiple Frequency/Time (MFT)” option (the required frequency can be selected during analysis).

4. Click on “Start”. Enter the file name and any comments you want to save. Collect data for 24-48 h.

5. As cells grow, C at high frequency (32 or 64 kHz) decreases (see Note 6 for comments on the choice of frequency). Confluence is achieved when C is stable at the minimum. At this point, the measurement can be stopped.

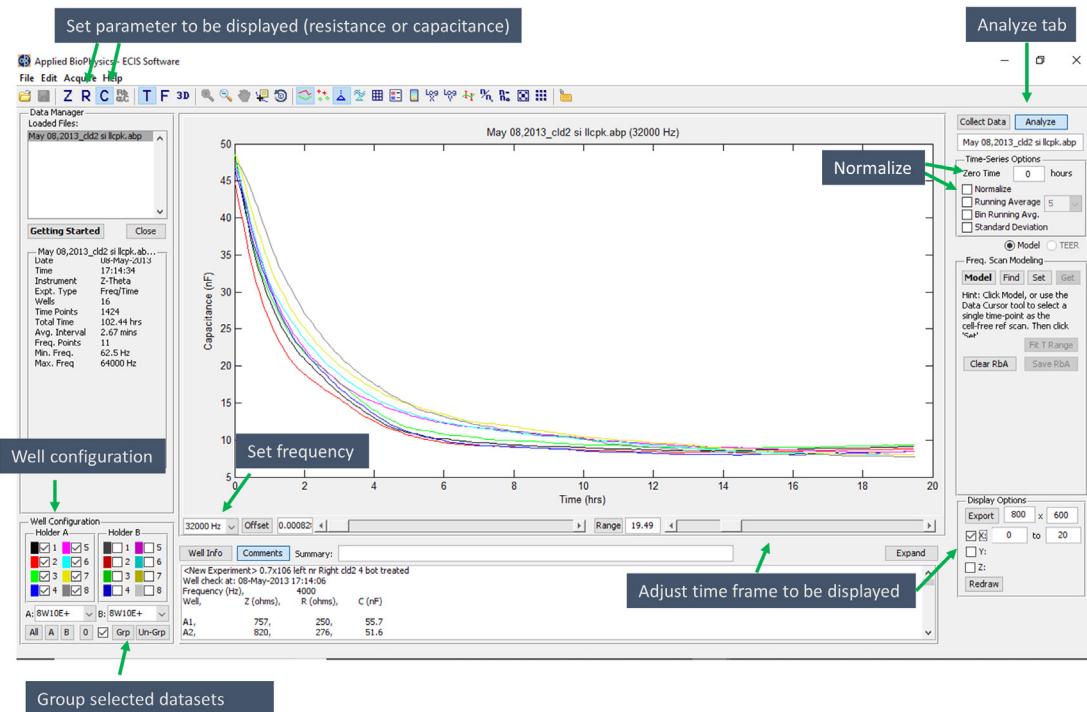
### **Data analysis**

1. Prepare data for analysis

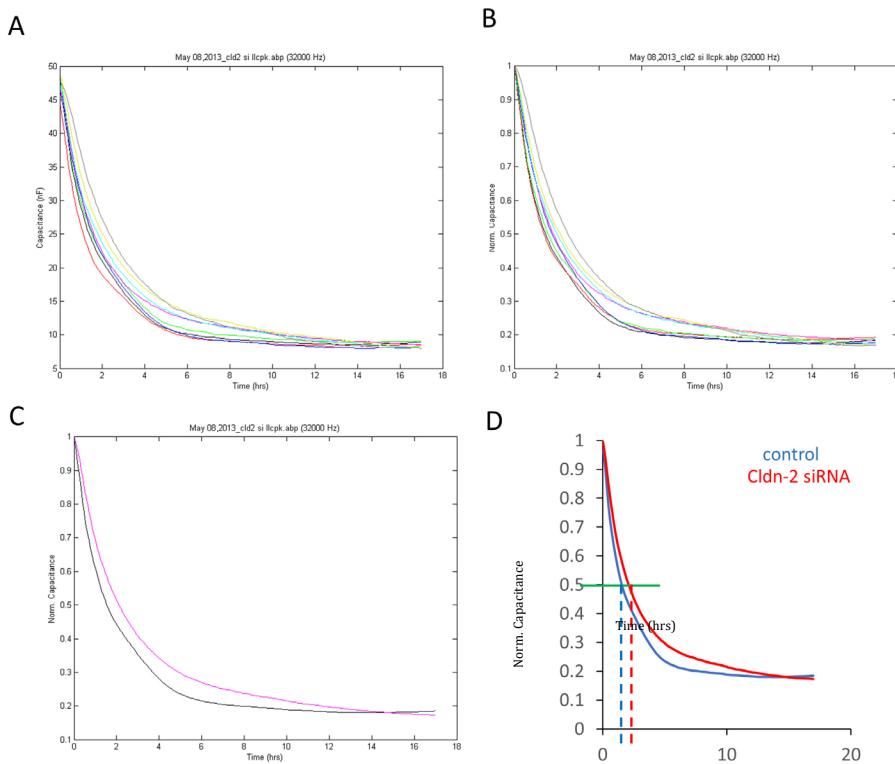
In the ECIS software on the right top click the “Analyze” tab. Figure 2 shows a screenshot of the software with a typical measurement, with the corresponding commands labelled. You can select the parameters and curves you want to export. This limits the amount of data exported.

Alternatively, you can export all data (see below) and process them in the graph software. However, selecting the data needed for your analysis prior exporting makes managing the dataset easier.

- a. **Select display parameters:** In “Well configuration,” select the wells you want to work with (Figure 2). On the top, select to display the parameter you would like to use and under the graph select to display the frequency. For cell growth, use capacitance (C) at a high frequency (32 or 64 kHz). For transepithelial resistance, select Z at a low frequency (we use 500 Hz).
- b. **Normalize data:** This can be useful for comparing measurements from different days, or if the variability of the starting values makes comparisons difficult. In the “Time series” option, click “Normalize”. Use the 0 h point for normalization. This will ensure that data collected on different days can be compared. Figures 3A and 3B show a measurement before and after normalization.
- c. **Combine parallel measurements:** You have the option to combine all parallel measurements and export them as a single dataset, to limit the size of the exported dataset. For this in “Well Configuration” select the wells that represent parallel measurements. Click on Grp (group). This will calculate the average of your measurements and display them as one curve. Calculate the average for each condition (Figure 3C).
- d. Make any other adjustment for exporting data: Adjust the graph to reflect the data you want to export: In “Well configuration” (bottom left), select the wells to export (either single measurements or the grouped value of the different conditions). Under the graph, adjust the X-axis to display the required time frame (Figure 2).



**Figure 2. Screenshot from the ECIS software displaying results from a typical measurement described in the protocol.** The cells were grown in an 8W10E+ array, placed in Holder A. Conditions were as follows: 1-4 (left side): NR siRNA transfected; 5-8 (right side) Cldn-2 siRNA transfected. The capacitance displayed is set to 32 kHz. The various options in the software described in the protocol are marked with green arrows and blue text boxes indicate the various options.



**Figure 3. Data exported using the ECIS software with or without various processing.** A-C. Exported graphs with various steps of analysis, as described in the protocol. A. All data are shown (graph from Figure 1). B. Data were normalized to the first point of the measurement. C. Data grouped according to the conditions. Black: NR siRNA; pink: Claudin-2 siRNA. D. Determination of half recovery time. Data were exported to Excel and graphed using a scatter graph with line option. Blue: NR siRNA; red: Claudin-2 siRNA. The green horizontal line indicates 0.5 C value (Y axis), and the blue and red dashed lines indicate the corresponding time (X axis value).

## 2. Exporting graph image and data

The graph can be exported as an image using the “File”, “Export graph” command. Once the image to be exported opens, chose the save icon on top, and set the file name and format you wish to use. This image however will be of low quality with small fonts. Another option is to export the data from the graph and process using a graph software. For this open “File”, select “Export data” and choose “Graph data”. This mode of exporting will only save the data displayed on the graph in a \*.csv format. You can also export all the data with the corresponding option in the “Export data” command. This will result in exporting all data collected at all frequencies.

## 3. Processing using a graphing software

Although the graph image can be exported, and the resulting picture has all the labels, these cannot be altered, and the graph and labels are of low quality. Therefore, we usually export the data to a graphing program for further processing and for generating publication quality graphs (Figure 3D).

#### 4. Analyzing TER

One mode of comparing barriers in two conditions is to compare  $Z$  at a given timepoint after junctions developed. In Figure 1B we graphed the  $Z$  values from two measurements (those shown in Figure 1A bottom graph and a similar series of experiment from a different day).

#### 5. Analyzing cell growth

To compare data obtained in different experiments, we calculate for each measurement the time required for the cells to cover half the electrode (the time corresponding to 0.5 C value). For this analysis we use the data from the capacitance (C) values at 32 kHz, normalized to the first measurement point (see step 2) (Figure 3D). The time values for all conditions are collected, and the mean  $\pm$  S.D. for each condition is calculated.

#### 6. Statistics

The measurements must be performed at least in 3 different experiments. We use at least duplicates for every condition on every array. Direct comparison between arrays can pose a challenge, as the raw values show variability in different experiments. For example, TER data combined from two arrays obtained on different days shown on Figure 1B reveal that data from the same experiment cluster closer together, but the values differ from those measured on another array. While in both cases the claudin-2 siRNA transfected cells show elevated TER, unpaired *t*-test on the combined dataset shows that the change is not significant. Normalizing and expressing data as fold changes can solve this problem, as described for the C values. If that method is chosen, a non-parametric statistical test should be used.

### Notes

1. **Array type:** There are various types of electrode arrays available, that differ in the number of electrodes. The more electrodes an array has, the more cells are being measured and averaged, resulting in reduced noise, but also smaller sensitivity. It is important to choose the appropriate electrode based on the experiment type. For a detailed description of each of the ECIS cultureware, users are referred to the Applied BioPhysics website: <https://www.biophysics.com/cultureware.php>.

For detecting cell growth and junction development, we used both 8W10E or 8W10E+. The 10E+ has more electrodes to collect information. Importantly, for a given series of experiment, always use the same array.

2. **Electrode coating:** Depending on the properties of the cell type used, collagen, fibronectin, laminin or other extracellular matrix proteins can be used to coat the electrode prior to seeding cells. Extracellular matrix proteins aid in cell adhesion and regulate a wide range of cell functions such as cell proliferation, differentiation, migration and survival. The type of protein used for coating might also affect proliferation and the development of the barrier, although this has to be systematically explored for specific cells. Therefore, it is important to standardize conditions of coating across experiments. We found that LLC-PK<sub>1</sub> cells do not require special coating, as

they grow well on the gold electrodes.

3. **Cell number and seeding:** The inoculation technique is key for comparability and reproducibility between wells. Difference in proliferation can be masked by a high variability between wells. This usually results from inconsistency of seeding, and variation in cell number. For accurate measurements, cell numbers must be kept constant across the wells. Preparing an even cell suspension for seeding is key. Thoroughly trypsinize cells and mix suspension to eliminate clusters of cells. Accurate counting of the cell suspension to obtain comparable numbers when using cells from different conditions (e.g., transfection) is also important. Finally, prewarming of the array and the cell suspension to eliminate temperature difference between the well bottom and the suspension is also vital. For following cell growth, low cell number must be used to allow the cells to proliferate after adhering. If the cell number is too high, the cells will be confluent after adhering and no proliferation will occur. Cell adhesion and spreading is usually complete in the first 3 h, and after this time frame the cells establish the confluent layer via proliferation.
4. Mycoplasma contamination of the cell lines should be routinely tested.
5. **Transfection parameters:** Transfection conditions vary depending on the cell lines used and the target proteins, therefore, optimization of the conditions may be required. Knockdown efficiency of the target protein(s) should be confirmed by western blotting or qPCR in parallel. Changes in confluence in the transfected cells should be assessed using the capacitance measurement. The cells on the arrays can be visualized using a microscopy. In addition, effects on individual tight junction proteins are also informative and can be assessed by immunofluorescence.
6. **Choice of frequency for capacitance and impedance measurement:** When setting up the system, it is recommended to use the setting that collects data at all available frequencies. Due to differences in the resistance against the flow of current through the cells and between the cells, parameters measured at different AC frequencies will reflect different aspects of cell properties. During analysis, the user can choose the frequency for the parameters of interest. Capacitance determined at high frequency reflects the coverage of the electrode, with minimal influence by other parameters, such as barrier function. The current recommendation and default in the software for analysis of capacitance values is 64 kHz. Of note, an earlier version of the software had 32 kHz as the highest and default C value, and therefore we used this setting in our earlier studies. At this frequency, C also reflects electrode coverage. In contrast, resistance of the paracellular space (referred to as barrier function, generated by the tight junctions) is best reflected by low AC frequencies. Impedance (Z) values at 500 Hz and below reflect dominantly resistance of the paracellular space. The software is also able to perform a mathematical modelling that allows further analysis and better distinction between resistance of the paracellular space and the space underneath the cells (cell-substrate attachment). For details the author is referred to Stolwijk *et al.* (2015).

## Recipes

1. 10 mM L-cysteine solution for electrode stabilization
  - a. Add 0.06 g of L-Cysteine to 50 ml dH<sub>2</sub>O
  - b. Filter to sterilize
  - c. Cysteine can easily oxidize when dissolved in water and therefore, this solution must be prepared fresh

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This protocol was derived from: Amoozadeh *et al.*, 2015; Dan *et al.*, 2019.

## Competing interests

The authors declare that they have no financial or non-financial conflicts of interest.

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