

Glioma Induction by Intracerebral Retrovirus Injection

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[Abstract] Glioblastoma (GBM) is the most common primary brain cancer in adults and has a poor prognosis. It is characterized by a high degree of cellular infiltration that leads to tumor recurrence, atypical hyperplasia, necrosis, and angiogenesis. Despite aggressive treatment modalities, current therapies are ineffective for GBM. Mouse GBM models not only provide a better understanding in the mechanisms of gliomagenesis, but also facilitate the drug discovery for treating this deadly cancer. A retroviral vector system that expresses PDGFBB (Platelet-derived growth factor BB) and inactivates PTEN (Phosphatase and tensin homolog) and P53 tumor suppressors provides a rapid and efficient induction of glioma in mice with full penetrance. In this protocol, we describe a simple and practical method for inducing GBM formation by retrovirus injection in the murine brain. This system gives a spatial and temporal control over the induction of glioma and allows the assessment of therapeutic effects with a bioluminescent reporter.

Keywords: Glioma, Retrovirus, Intracerebral injection, Murine model, PDGFBB, PTEN, P53

[Background] Glioblastoma (GBM) is the most aggressive malignant brain tumor and unfortunately is also almost always fatal. Despite advances in multiple therapeutic modalities, no effective therapy has been developed to cure GBM. The mechanisms underlying this disease remain poorly understood. Animal models have been a very important tool to define the GBM pathogenesis and test for gene or drug therapeutics. Several mouse models have been developed with the aim to produce a disease which mimics the human disease as closely as possible and exhibits similar molecular, genetic and histological character. The prominently used models are xenograft (Hingtgen *et al.*, 2008) models where human tumor cell lines can be transplanted orthotopically in brain as well as ectopically in subcutaneous area in immunocompromised mice, providing an advantage of having a large tumor mass in a brief period. Genetically engineered mice models (GEMM) with specific gain-of oncogenic activities or loss of tumor suppressor pathways, which resemble perturbations in the most frequently dysregulated pathways in GBM, results in formation of gliomas in rodents. Genetic perturbations in GEMM models often include the gain-of-function mutations in oncogenic factors such as EGFR, PI-3K, and Ras (Holland *et al.*, 2000; Zhu *et al.*, 2009), and PDGF amplification (Assanah *et al.*, 2006), as well as the loss-of-function in tumor suppressors such as NF1 (Zhu *et al.*, 2005), TP53 (p53) (Zheng *et al.*, 2008), Ink4a/ARF (Holland, 2001), PTEN (Holland *et al.*, 1998 and 2000). The use of viral vectors (Ikawa *et al.*, 2003; Hambardzumyan *et al.*, 2009; Marumoto *et al.*, 2009; Friedmann-Morvinski *et al.*, 2012) such as retrovirus, lentivirus and adenovirus carrying oncogenes and/or targeting tumor suppressors has been a convenient and effective

approach to induce brain tumor formation in rodents. There is a need for testing multiple different tumor models for therapeutics since different subtypes of gliomas have varied genetic profiles and clinical responses to drug treatment.

GBM have been classified into four subtypes depending upon their gene expression pattern and molecular signature, namely, Mesenchymal, Neural, Proneural and Classical tumors (Verhaak *et al.*, 2010). The Proneural subtype of GBM predominantly involves mutations/loss in oncogene P53 and amplification of PDGFR α with loss of PTEN seen across all the subtypes (Verhaak *et al.*, 2010; Lei *et al.*, 2011). To closely imitate the human Proneural subtype, Lei *et al.* (2011) devised a GBM model by inducing *PTEN* and *P53* deletion and *PDGFBB* overexpression in the progenitor cells of the white matter region of adult murine brain through retrovirus inoculation (Lei *et al.*, 2011). We have successfully used this retrovirus-induced GBM model in mice ranging in age from day 2 to 3-month-old adult mice with remarkable reproducibility (Lu *et al.*, 2016). This model develops gliomas with full penetrance within 3-4 weeks which in contrast to other GEMM models, which take a significantly longer time.

Materials and Reagents

A. Materials needed for culture and virus packaging

1. Culture dishes 10 cm diameter (Corning, Falcon[®], catalog number: 353003)
2. 0.45 μ m size filter (Corning[®] bottle-top vacuum filter system 500 ml) (Corning, catalog number: 430512)
3. 38 ml open-end tubes (Beckman Coulter, catalog number: 344058)
4. 50 ml screw capped tubes (Corning, Falcon[®], catalog number: 352098)
5. 15 ml screw capped conical tubes (Corning, Falcon[®], catalog number: 352196)
6. Round bottom 5 ml tubes (Beckman Coulter, catalog number: 344057)
7. Parafilm
8. Vector system
 - a. Retroviral vector carrying PDGFB-IRES-Cre
 - b. Retroviral packaging plasmid pEco (Clontech, catalog number: PT3749-5)

Note: We use a two-vector system for retrovirus production. One is a retroviral vector carrying PDGFB-IRES-Cre, which overexpresses PDGFB and Cre, and another is a retroviral packaging plasmid, which provides gag, pol and env to produce VSV-G pseudotyped retrovirus. We use commercially available DNA transfection reagents for 293T HEK transfection to produce the retrovirus.

9. Low passaged 293T HEK cells (P6-P14)
10. Dulbecco's modified Eagle medium (DMEM) (Thermo Fisher Scientific, GibcoTM, catalog number: 11960044)
11. Fetal bovine serum (FBS) (Atlanta Biologicals, catalog number: S11550)
12. L-glutamate (Thermo Fisher Scientific, GibcoTM, catalog number: 25030081)
13. Penicillin-streptomycin (Thermo Fisher Scientific, GibcoTM, catalog number: 15140122)

14. Sodium pyruvate (Thermo Fisher Scientific, GibcoTM, catalog number: 11360070)
15. Trypsin-EDTA (Thermo Fisher Scientific, GibcoTM, catalog number: 25300054)
16. Transfection agent: Polyjet (SignaGen Laboratories, catalog number: SL100688)
17. 20% sucrose in HBSS (Hank's Balanced Salt Solution) (Thermo Fisher Scientific, GibcoTM, catalog number: 14025134)
18. Complete media for 293T cells (D10 culture media) (see Recipes)

B. Materials needed for titration of retrovirus

1. 24-well plate
2. Isolated MEF cells (10^5 cells/well)
3. CAG-Rosa-tdTomato reporter mice (THE JACKSON LABORATORY, catalog number: 007909)
4. Concentrated virus
5. Poly-L-lysine (Sigma-Aldrich, catalog number: P7890), 0.05 mg/ml prepared in sterile tissue-culture grade water, filter sterilized
6. Sterile tissue culture grade water (Sigma-Aldrich, catalog number: W3500)
7. Dulbecco's modified Eagle medium (DMEM) (Thermo Fisher Scientific, GibcoTM, catalog number: 11960044)
8. Fetal bovine serum (FBS) (Atlas Biologicals, catalog number: S11550)
9. L-glutamate (Thermo Fisher Scientific, GibcoTM, catalog number: 25030081)
10. Penicillin-streptomycin (Thermo Fisher Scientific, GibcoTM, catalog number: 15140122)
11. Sodium pyruvate (Thermo Fisher Scientific, GibcoTM, catalog number: 11360070)
12. Complete media for MEF cells (D10 culture media) (see Recipes)

C. Materials needed for intracranial stereotaxic injection

1. 1 cc syringes (BD, catalog number: 309659) and 30 G needles (BD, catalog number: 305128)
2. Autoclaved cotton tips
3. Animals

Pten^{f/f} (THE JACKSON LABORATORY, catalog number: 006440)

Trp53^{f/f} (THE JACKSON LABORATORY, catalog number: 008462)

Notes:

- a. *These mouse stains were crossed with a reporter line Rosa-tmLuciferase (THE JACKSON LABORATORY, catalog number: 005125) which has a firefly luciferase gene inserted at the ROSA26 locus (Durkin et al., 2013). This bioluminescent reporter will help to track the tumor growth and evaluate candidate therapeutic regimens. The mice were maintained on a mixed C57Bl/6; 129Sv; CD-1 background.*
- b. *Obtain appropriate animal use protocol from Institutional Animal Care and Use Committee (IACUC) before the animal work.*

4. Polybrene (1 μ g/ml) (Sigma-Aldrich, catalog number: 107689)

5. Isoflurane (Piramal[®]) for inhalational anesthesia (2-3% for induction), or Intraperitoneal Route: ketamine (Ketaset[®], NDC 0856-2013-01) + xylazine (Anased[®] 20 mg/ml, Santa Cruz Biotechnology, catalog number: sc-362950Rx) combination
6. Buprenorphine (Buprenex[®] 0.3 mg/ml): Dilution 1 ml of Buprenex + 9.0 ml D5W for use in mice
7. Sterile phosphate buffered saline
8. 70% ethanol
9. Chlorhexidine (BactoShield[®] CHG 2% Surgical Scrub) (STERIS, catalog number: 132224)
10. Isopropanol (Priority Care[®], Isopropyl Alcohol 70%) (FIRST PRIORITY, catalog number: MS070PC)
11. GLUture[®] Topical Tissue Adhesive (Abbott, catalog number: 32046-04-01)
12. Artificial Tears Lubricant Ophthalmic Ointment (Henry Schein, catalog number: 18581)
13. Ketamine and xylazine combination (see Recipes)

Equipment

- A. Equipment needed for ultracentrifugation concentration
 1. Pipettes
 2. Sterile working hood
 3. Ultracentrifuge (Beckman Coulter, model: Optima XPN-90 or equivalent)
 4. Shaker
 5. Rotors:
 - a. SW 32 Ti (Beckman Coulter, catalog number: 369650)
 - b. SW 55 Ti (Beckman Coulter, catalog number: 342196)
- B. Equipment needed for titration of retrovirus
 1. Biosafety hood
 2. Neubauer cell counting chamber
- C. Equipment needed for surgery
 1. Stereotaxic device (RWD Life Science, catalog number: 68502 with mouse adapter: RWD Life Science, catalog number: 68010)
 2. Micro-drill (RWD Life Science, catalog number: 78001) to make a burr hole in the skull with appropriate drill bits (RWD Life Science, catalog number: 78002)
 3. Autoclaved surgical pack composed of scissors, forceps, cotton swabs and tips, applicators, scalpel blade holder
 4. 10 μ l Hamilton syringe (Hamilton, catalog number: 7659-01) with 30 G blunt-end needle (Hamilton, catalog number: 7803-07)
 5. Incubator or heating pad
 6. Timer

7. Biobubble or sterile working place
8. Surgical site Marker (MEDTRONIC, Devon™, catalog number: 31145793)
9. Anesthesia Machine (Ohmeda, model: Excel 210SE or equivalent)

Procedure

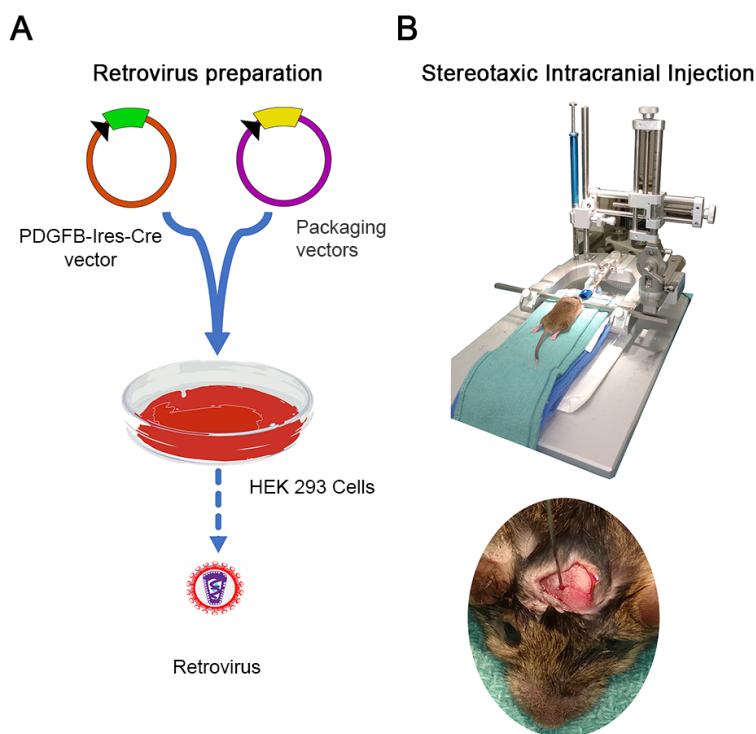


Figure 1. Schematic of the whole procedure. A. Viral packaging and concentration; B. Stereotaxic intracranial injection of virus for induction of glioma.

A. Transfection and viral packaging procedure (Figure 1A)

Note: We follow the manufacturer provided protocol for Polyjet mediated transfection (<http://signagen.com/DataSheet/SL100688.pdf>).

1. Day 1: Plating of 293T cells

Plate 293T cells (around $3-4 \times 10^6$) in poly-L-lysine coated 10 cm diameter dishes in DMEM with 10% FBS media (D10 culture media). Cells will become 70-80% confluent after 24 h.

Note: Plan accordingly to allow 5-6 h next day for transfection and medium change.

2. Day 2: Transfection day

a. Pre-transfection check

- i. Check for confluence next day and use the plates with 70-80% cell confluence.

Note: Higher confluence can be a hindrance as cells don't have space to grow.

- ii. 1 h before transfection, change media with fresh D10 media—4 ml per plate.

- iii. Use the following table for 10 cm plates for transfection (Table 1).

Table 1. Transfection mix

For 10 cm plate	
Vector DNA (PDGFB-IRES-Cre)	7.5 µg
Packaging plasmid	7.5 µg
Polyjet (3x total DNA content)	45 µl in 455 µl FBS free DMEM
D10 culture media	4 ml
Total transfection volume	5ml

b. Transfection

- Mix the calculated concentration of DNA in 500 µl DMEM without FBS and gently mix.
- Add 45 µl of Polyjet in 455 µl of serum free DMEM media and gently mix with pipette and immediately mix with diluted DNA of step A2b i. Gently mix them and incubate at room temperature for 10-15 min.
Note: Not more than 20 min.
- Then add it into the plates drop wise and gently shake.
- Incubate at 37 °C and 5% CO₂ for 5 h.
Note: Do not keep longer as it can be toxic to the cells.
- Discard the transfecting media and add fresh D10 10 ml per plate and incubate at 37 °C and 5% CO₂ for 24 h.

3. Day 3: 1st collection

- Collect the media and centrifuge at 600 x g (2,000 rpm) for 10 min and collect supernatant, leaving the cellular debris at the bottom. Collected media can be stored at 4 °C for 3 days at max (Tiscornia *et al.*, 2006) before ultracentrifuge concentration.
- Add fresh D10 media 10 ml per plate and incubate again for next 24 h.

4. Day 4: 2nd collection

- Collect the media and centrifuge at 600 x g (2,000 rpm) for 10 min and collect the supernatant, leaving the cellular debris at the bottom.
- Pool the media from day 3 and day 4 and filter using 0.45 µm pore size filter.

5. Ultracentrifugation for viral concentration

- We use Beckman Coulter rotor SW 32 Ti, which has capacity to hold six 38 ml open-end tubes. Balance the tubes to the maximum difference of 0.5 g and centrifuge at 68,300 x g (20,000 rpm) for 2 h at 4 °C. With each tube holding 30 ml, a total 360 ml of media can be centrifuged in one run. If required to run more media, discard the supernatant after 1st spin without disturbing the pellet add fresh media for 2nd run.
Note: Do not use the tubes filled less than half of its capacity which increases the risk of collapse.
- Pour off the supernatant after the run and aspirate rest of the droplets from the wall without disturbing the pellet.

- c. Transfer the centrifuge tubes in to 50 ml screw capped tubes and keep them upright on shaker at 4 °C overnight and allow the residual media to re-suspend pellet (roughly around 20-30 µl/tube).
- d. Centrifuge briefly and collect the re-suspended pellets from all the tubes, aliquot the concentrate in 4-10 µl volumes and store at -80 °C for more than a month. Avoid repeated cycles of thawing and freezing that can drastically reduce the viral titer.
- e. For *in vivo* injection, further purification of virus by sucrose cushion is recommended (Tiscornia *et al.*, 2006).

6. Sucrose cushion
 - a. Collect the viral concentrate after overnight shaking and wash the tube with 100 µl of HBSS and pool it with collected viral concentrate.
 - b. Using round bottom 5 ml tubes, fill it with 1.5 ml of 20% sucrose in HBSS and add 200 µl of viral concentrate. The rotor used is SW 55 Ti and after balancing, spin at 48,600 *x g* (20,000 rpm) for 2 h at 4 °C.
 - c. Pour off the supernatant and allow rest to drain off by inverting the tubes. Re-suspend the pellet in 50-100 µl of HBSS, cover with Parafilm and shake slowly for 20-30 min.
 - d. Collect the concentrate, aliquot it and store at -80 °C.
 - e. Check for the titer in a reporter cell line. We use mouse embryonal fibroblasts, which express tdTomato reporter upon Cre expression.

B. Titration of retrovirus

MEF (mouse embryonic fibroblasts) cells are isolated from E13.5-14.5 embryos of CAG-Rosa-tdTomato reporter mice (THE JACKSON LABORATORY) (Durkin *et al.*, 2013) for retrovirus titering. These cells express tdTomato when exposed to Cre-expressing viruses and allowing us to assess the viral infectivity. For virus titration, we use the procedure described by Tiscornia *et al.* (2006).

1. Coat the 24-well plate with 500 µl poly-L-lysine per well with incubation at 37 °C for 30 min, aspirate the media and wash with autoclaved deionized water and let it dry for 1-2 h in biosafety hood.
2. Plate 10⁵ MEF cells in 500 µl D10 media per well in 24-well plate. Use a minimum of 8 wells including one as negative control.
3. Incubate it for 6 h and then check after 6 h if they adhere to the plate.
4. Make 10-fold dilutions of retrovirus (undiluted to 10⁶) in PBS with total volume around 50 µl.
5. Change 480 µl D10 media with polybrene at a concentration of 4 µg/ml.
6. Add 20 µl of each viral dilution to the well and mix gently and thoroughly.
7. Incubate the cells at 37 °C for 48 h. TdTomato will be visible within 24-48 h.
8. Assess the percentage of tdTomato positive cells by counting on Neubauer cell counting chamber or by using FACS and calculate the titer by using following formula:

$$\text{Biologically Active Viral Particles/ml} = \frac{\text{Number of infected cells (tdTomato+)} \times \text{Dilution Factor}}{\text{Volume of Virus in ml}}$$

Viral titer $\geq 10^7$ /ml is adequate for tumor formation.

C. Intracranial stereotaxic injection procedure (Figure 1B)

Note: Neonatal or young adult mice can also be used with appropriate stereotaxic device adapter.

1. Pre-surgical preparation

- a. Switch on the incubator or heating pad.
- b. Check the micro-drill, anesthesia machine for oxygen pressure and adequate isoflurane flow and overhead light source.
- c. Set up the stereotaxic instrument as per the instructions of the manufacturer.
- d. Switch on the anesthesia machine with attachment for mouse enclosure. Oxygen flow (0.8-1.5 L/min) and isoflurane (2-3%) are set for induction of anesthesia and decreased to 1-1.5% for maintenance. Alternatively, use ketamine and xylazine combination which is injected intra-peritoneal.
- e. Load the concentrated virus in syringe (e.g., 3 μ l per each mouse to be injected) slowly avoiding bubbles and set it up on the holder in stereotaxic device.
- f. Apply artificial tear ointment on exposed cornea to avoid drying.

2. Surgical procedure

- a. Anesthetize the mouse and check the anesthesia level by toe pinch.
- b. Set up the mice on stereotaxic device with nose and ear holders firmly to avoid any accessory movement of head during the procedure and isolate the surgical area with sterile drape or covers.
- c. Clean the operative area (back of the skull) using autoclaved cotton tips with chlorhexidine and isopropanol alternatively three times each. Wait for a few minutes.
- d. Make an incision on the back of head of about 1-1.5 cm in length to expose bregma.
- e. Adjust the stereotaxic device in relation to bregma as a reference and then use the coordinates depending on the age and area of brain to be targeted (e.g., for targeting the corpus callosum in adult mice, we used the coordinates: Anteroposterior + 0.2 mm, Lateral ± 0.5 mm and dorso-ventral 2.2 mm).
- f. Mark the targeted area with surgical site marker.
- g. Drill hole in bone using micro-drill being careful to avoid going too deep.
- h. Lower the syringe needle 0.4-0.6 mm deeper to the desired depth (e.g., here 2.2 + 0.6 mm), wait for 1-2 min and retract back to desired depth to form a small pocket and slowly inject 1 μ l of viral concentrate, wait for 1-2 min, retract 1 mm and inject 1 μ l again. We usually inject 1-3 μ l of viral concentrate in one adult mouse.
- i. Slowly retract the needle to avoid leaking of the concentrate.
- j. After complete retraction, use tissue glue to join the tissue flaps.
- k. Give buprenorphine injection intra-peritoneal. (0.05-0.1 mg/kg)
- l. Keep the mouse on heating pad or incubator till it becomes conscious.
- m. Monitor the mice for next 2-3 days and administer analgesics as per the protocol.

- n. Wash the needle with 70% ethanol and three times with sterile water.

Data analysis

The initial tumor growth can be noticed within 2-3 weeks in the brain monitored by *in vivo* bioluminescent imaging system such as IVIS Spectrum CT (PerkinElmer). Tumor-bearing mice normally survive for 3-6 weeks post injection depending on age at the time of injection. Neonatal mice injected at age postnatal day 2 (P2) to day 4 (P4) (48-96 h) tend to survive longer and develop a larger tumor, possibly due to the expansion of non-fused calvarial bones. The brain sections are stained with hematoxylin and eosin and tumors observed are typically invasive and hypercellular with hyperchromatic nuclei, foci of palisading necrosis and microvascular proliferation suggestive of glioblastoma (Figure 2).

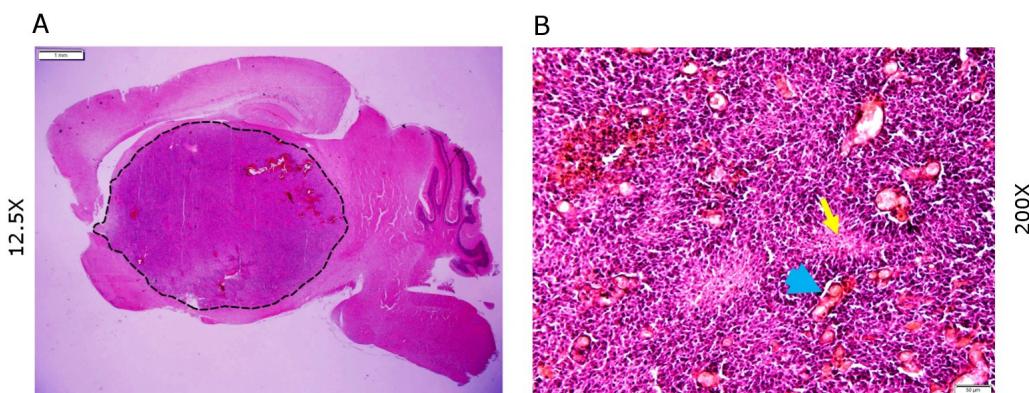


Figure 2. H&E histology of tumor sections. A. Sections showing tumor growth in the brain (Dotted line enclosure); B. Section showing typical features of glioblastoma-palisading necrosis (yellow arrow) and microvascular proliferation (blue arrowheads). Scale bars in (A) 1 mm and (B) 50 μ m.

Notes

1. Use only healthy and low passaged 293T or 293FT cells for virus packaging.
2. Do not use filter smaller than 0.45 μ m for the risk of shearing effect on viral particles.

Recipes

1. Complete media for 293T and MEF cells (D10 culture media)
435 ml DMEM medium supplemented with 50 ml FBS
5 ml L-glutamine
5 ml penicillin-streptomycin
5 ml sodium pyruvate

2. Ketamine and xylazine combination
1 ml ketamine (Ketaset® 100 mg/ml)
0.5 ml of xylazine (Anased® 20 mg/ml)
8.5 ml of D5W (5% dextrose in water)

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