

Rapid Isolation of Total Protein from *Arabidopsis* Pollen

Ming Chang^{1,2} and Shanjin Huang^{1,2,*}

¹Center for Plant Biology, School of Life Sciences, Tsinghua University, Beijing 100084, China; ²Key Laboratory of Plant Molecular Physiology, Institute of Botany, Chinese Academy of Sciences, Beijing 100093, China

*For correspondence: sjhuang@tsinghua.edu.cn

[Abstract] *Arabidopsis* pollen is an excellent system for answering important biological questions about the establishment and maintenance of cellular polarity and polar cell growth, because these processes are amenable to the genetic and genomic approaches that are readily available in *Arabidopsis*. Given that proteins are the direct executors of a wide variety of cellular processes, it is important to rapidly and efficiently isolate total protein for various protein-based analyses, such as Western blotting, co-immunoprecipitation and mass spectrometry, among others. Here we present a protocol for rapid isolation of total protein from *Arabidopsis* pollen, which is adapted from our recently published paper (Chang and Huang, 2015).

Keywords: *Arabidopsis thaliana*, Flower, Pollen, Protein isolation, Western blot

[Background] Pollen is a critical stage during the life cycle of sexually-reproducing plants. Pollen germination and subsequent tube growth provide the passage for two non-motile sperm cells to effect double fertilization in flowering plants. Pollen is routinely used as a model system to addressing fundamental cell biological questions, such as the establishment and maintenance of cell polarity and polar cell growth, as well as the structure and function of the actin cytoskeleton (Chen *et al.*, 2009; Qu *et al.*, 2015). Generally, pollen derived from lily and tobacco was widely used due to the large size of the grains, which makes them easy to collect and observe under the microscope. In contrast, *Arabidopsis* pollen is small and it is relatively difficult to collect a large amount of pollen to isolate a sufficient quantity of protein for downstream analyses such as Western blotting and mass spectrometry. Therefore, development of a protocol to rapidly isolate total protein from *Arabidopsis* pollen will facilitate related analyses.

Materials and Reagents

1. 1.5 ml Eppendorf tubes (Fisher Scientific, FisherbrandTM, catalog number: 05-408-129)
2. Pipette tips (USA Scientific, catalog numbers: 1112-1720, 1110-1200)
3. *Arabidopsis* plants (ecotype Col-0)
4. Ponceau S (Sigma-Aldrich, catalog number: P3504)
5. Anti- α -tubulin (Beyotime Biotechnology, catalog number: AT819)
6. Anti-actin (Abmart, catalog number: M20009)

7. Anti-COXII (Agrisera, catalog number: AS04 053A)
8. 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) (AMRESCO, catalog number: 0511)
9. K-acetate (Sinopharm Chemical Reagents, catalog number: 30154518)
10. Magnesium chloride hexahydrate ($MgCl_2 \cdot 6H_2O$) (Sigma, catalog number: 63064)
11. Tween 20 (AMRESCO, catalog number: 0777)
12. Triton X-100 (Sigma-Aldrich, catalog number: V900502)
13. Phenylmethanesulfonyl fluoride (PMSF) (EMD Millipore, catalog number: 52332)
14. K-HEPES protein extraction buffer (see Recipes)

Equipment

1. Pestles (VWR, catalog number: 89093-446)
2. Forceps (Shanghai Haiou)
3. Pipettes (VWR, catalog numbers: 89079-970, 89079-974)
4. Vortex (Corning, model: Corning[®] LSETM Vortex Mixer)
5. Centrifuge (Eppendorf, model: 5417 R)

Procedure

1. Grow *Arabidopsis* plants in a growth room at 21 °C under long-day conditions (16 h light/8 h dark) for 6 to 8 weeks. Normally, the Col-0 ecotype will produce many flowers under these conditions. The plant growth time can be changed to adapt to the flowering time of other *Arabidopsis* ecotypes.
2. Quickly collect more than 100 freshly opened flowers (stage 13-15, Figure 1) in one 1.5 ml Eppendorf tube and add 1 ml K-HEPES protein extraction buffer (see Recipes). This step can be done at room temperature.

*Note: Collect *Arabidopsis* flowers around 10 AM. At this time the flowers will be fully opened, allowing easier access to the pollen.*



Figure 1. An opening *Arabidopsis* flower at stage 13-15 (Müller, 1961; Smyth *et al.*, 1990)

3. Vortex the flowers at the highest speed for 2-5 min in order to thoroughly separate the pollen from the flowers (Video 1).

Video 1. Procedure for the isolation of pollen from flowers by vortex in step 3



4. Centrifuge at 350 x g for 1 min at 4 °C (Video 2).

Note: Under low speed, most of the flowers will be suspended in the upper layer of buffer, but the pollen will be centrifuged to the bottom of the tube.

Video 2. Procedure for the isolation of protein as stated in step 4-7



5. Slowly and thoroughly remove all the flower debris with small forceps (Video 2).

Note: If necessary, you can repeat steps 4 and 5 several times for total removal of the flower debris, but you should be careful to avoid touching the pollen at the bottom of the tube.

6. Completely remove the supernatant (Video 2).

Note: Make sure the pollen is still at the bottom of the tube.

7. Put the tube on ice and thoroughly grind the pollen with a small pestle for 1-3 min (Video 2).

The liquid mixture will turn yellow color if the pollen is homogenized completely.

Note: If your lab doesn't have suitable pestles for 1.5 ml tubes, you can use a 1 ml pipette tip, sealed by melting it in a flame, as a substitute.

8. Add 50 μ l K-HEPES protein extraction buffer into the tube and keep the tube on ice for about 1 h.

9. Centrifuge at 18,000 \times g for 20 min at 4 °C, then transfer the supernatant to a new 1.5 ml tube (keep the tube on ice). Repeat this step twice. The supernatant is the final pollen protein sample and the protein concentration can be measured by standard Bradford assay.

Data analysis

SDS-PAGE and Western blot analyses of *Arabidopsis* pollen total proteins are shown in Figure 2. Total proteins isolated from *Arabidopsis* rosette leaves, in which the small subunit of Rubisco is by far the most abundant component, were used as the control (Figure 2A). In contrast, the band corresponding to the small subunit of Rubisco is hardly detected in the pollen total proteins (Figure 2B). This means that detection of proteins other than the small subunit of Rubisco might be easier in *Arabidopsis* total protein extract. For instance, by loading 10 μ g of total proteins from pollen per lane, α -tubulin (TUA), actin (ACT), cytochrome c oxidase subunit II (COXII) and profilin (PRF) can be easily detected (Figure 2B).

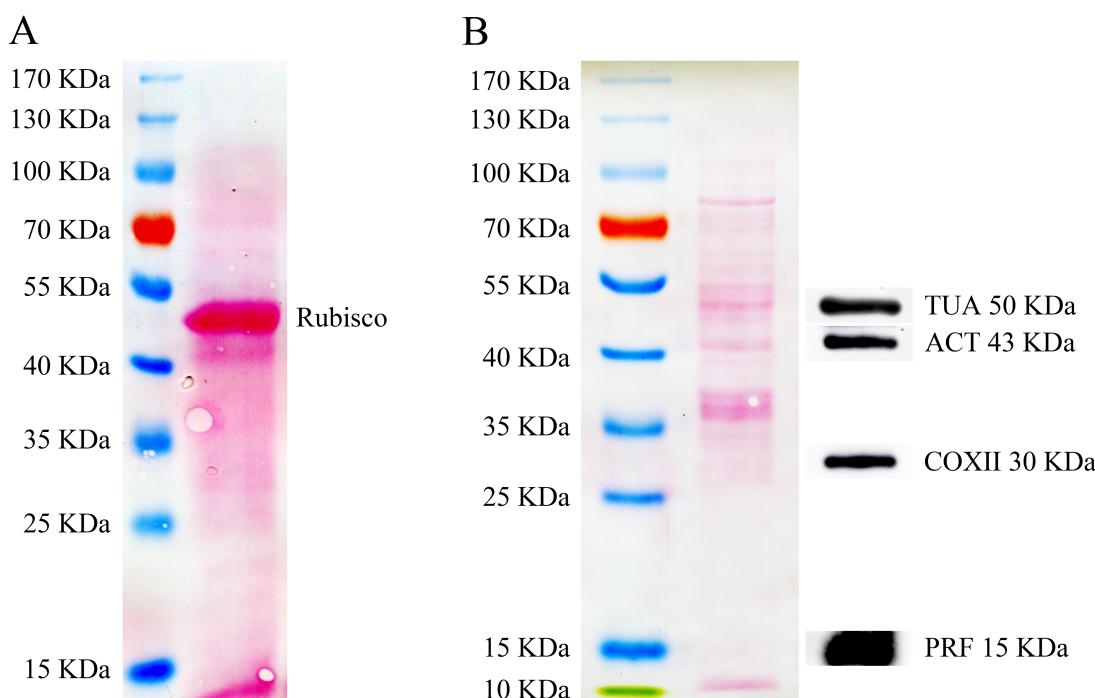


Figure 2. SDS-PAGE analysis of total proteins isolated from *Arabidopsis* pollen and leaves, and Western blot analysis of total proteins from *Arabidopsis* pollen probed with several antibodies. A. SDS-PAGE analysis of total proteins (50 µg) isolated from *Arabidopsis* rosette leaves. The nitrocellulose (NC) membrane was stained with Ponceau S after protein transfer. B. The left panel shows the SDS-PAGE analysis of total proteins (10 µg) isolated from *Arabidopsis* pollen. The NC membrane was stained with Ponceau S after protein transfer. The right panel shows the Western blot analysis of *Arabidopsis* pollen total proteins probed with anti- α -tubulin, anti-actin, anti-COXII and profilin antibodies. The profilin antibody was generated in our laboratory; see the details in our published paper (Liu *et al.*, 2015). Other antibodies are commercially available: anti- α -tubulin (Beyotime Biotechnology, AT819), anti-actin (Abmart, M20009) and anti-COXII (Agrisera, AS04 053A).

Notes

Generally, 100 Col-0 flowers will yield about 100 µg total protein from pollen with this method.

Recipes

1. K-HEPES protein extraction buffer
20 mM HEPES, pH 7.0
110 mM K-acetate
2 mM MgCl₂
0.1% Tween 20
0.2% Triton X-100

1 mM PMSF (add immediately before using)

Note: This buffer (without the PMSF) can be stored for up to 6 months at 4 °C.

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