

Extraction and Profiling of Plant Polar Glycerol Lipids

Yu Liu^{1, 2*} and Xuemin Wang^{1, 2}

¹Department of Biology, University of Missouri-St. Louis, St. Louis, USA; ²Donald Danforth Plant Science Center, St. Louis, USA

*For correspondence: yuliu@danforthcenter.org

[Abstract] This protocol describes a method to extract total polar glycerol lipids from plant materials, followed by mass spectrometry profiling. Different glycerol lipid classes can be distinguished by their head-groups, which can be profiled automatically and quantitatively by a triple quadrupole mass spectrometry in multiple reaction monitoring (MRM) mode with an autosampler. Comparing with other established methods, such as thin layer chromatography (TLC) separation followed by Gas spectrometry (GC) analysis, this method requires little effort in sample preparation and separation, while the resolution is not limited to general lipid classes but at side chain level. This method was described and used successfully to profile plant lipids changes under freezing stress in Welts *et al.* (2002).

Materials and Reagents

1. PYREXTM screw cap culture tubes with PTFE lined phenolic caps (Thermo Fisher Scientific, catalog number: 14-933D)
2. Disposable glass pipettes (Thermo Fisher Scientific, Fisher ScientificTM, catalog number: 13-678-20C)
3. Autosampler vials (Sigma-Aldrich, catalog number: 29133-U) with pre-cut cap (Sigma-Aldrich, catalog number: 29494-U)
4. TLC Silica gel 60 plate (Merck)
5. 1 ml syringe
6. Injection loop
7. 4-6 weeks old *Arabidopsis* rosette leaves
Note: Other plant materials can be used, additional precautions may be needed (see Notes).
8. Isopropanol (Thermo Fisher Scientific, catalog number: A416)
9. Potassium chloride (KCl) (Sigma-Aldrich, catalog number: P9333)
10. Chloroform (Thermo Fisher Scientific, catalog number: C607)
11. Methanol (Thermo Fisher Scientific, Fisher ScientificTM, catalog number: A456)

12. Acetic acid
13. Iodine
14. Primuline
15. Acetone
16. Nitrogen gas
17. Butylated hydroxytoluene (BHT) (Sigma-Aldrich, catalog number: B1378)
18. 7.5 M ammonium acetate solution (NH₄Ac) (Sigma-Aldrich, catalog number: A2706)
19. Phospholipid (PL) standard [kindly provided by Dr. Ruth Welti, (Kansas State University)]
Note: An example PL standard list is shown in Table 1.
20. Monogalactosyldiacylglycerol (MGDG), hydrogenated (Matreya, catalog number: 1058)
21. Digalactosyldiacylglycerol (DGDG), hydrogenated (Matreya, catalog number: 1059)
Note: The actual composition of MGDG and DGDG standards need to be determined experimentally, such as transmethylation followed by GC analysis (Politz et al., 2013).
22. Isopropanol solution with BHT (see Recipes)
23. Lipid extraction solution (see Recipes)
24. KCl solution (see Recipes)
25. Solution B (see Recipes)
26. Galactolipid (GL) standard (see Recipes)
27. Phospholipid (PL) standard (see Recipes)

Equipment

1. Water bath (Thermo Fisher Scientific, model: Isotemp2340)
2. Water bath shaker (Labnet International, model: SWB5050)
3. Nitrogen evaporator N-EVAP (Organomation, model: 111)
4. Fume hood
5. Ultraviolet light (360 nm)
6. Oven (Thermo Fisher Scientific, Fisher Scientific™, model: 13-247-650G)
7. API 4000 LC-MS/MS system (AB SCIEX) with Turbolonspray probe and HTC PAL autosampler (LEAP Technologies)

Table 1. Example of PL standard list

PL species	Mass of ion	nmol/sample	Used to quantify	Source
PG di 14:0	684.5	0.30	PG	Avanti Lipids Polar, catalog number: 840445
PG di Phy (20:0)	852.7	0.30		Avanti Lipids Polar, catalog number: 850404
LysoPE 14:0	426.3	0.30	LysoPE	Avanti Lipids Polar, catalog number: 856735
LysoPE 18:0	482.3	0.30		Avanti Lipids Polar, catalog number: 856715
PE di 12:0	580.5	0.30	PE	Avanti Lipids Polar, catalog number: 850702
PE di Phy (20:0)	804.6	0.30		Avanti Lipids Polar, catalog number: 850402
LysoPC 13:0	454.3	0.60	LysoPC	Avanti Lipids Polar, catalog number: 855476
LysoPC 19:0	538.3	0.60		Avanti Lipids Polar, catalog number: 855776
PC di 12:0	622.4	0.60	PC	Avanti Lipids Polar, catalog number: 850335
PC di 24:1	954.9	0.60		Avanti Lipids Polar, catalog number: 850399
PA di 14:0	400.3	0.30	PA	Avanti Lipids Polar, catalog number: 830845
PA di Phy (20:0)	456.3	0.30		Avanti Lipids Polar, catalog number: 850406
PS di 14:0	680.6	0.20	PS	Avanti Lipids Polar, catalog number: 840033
PS di Phy (20:0)	848.8	0.20		Avanti Lipids Polar, catalog number: 850408
PI 16:0-18:0	856.7	0.234	PI	Avanti Lipids Polar, custom synthesis
PI di 18:0	884.7	0.20		Avanti Lipids Polar, catalog number: 850143

PE: phosphatidylethanolamine; LYSOPE: lyso-phosphatidylethanolamine; PC: phosphatidylcholine; LYSOPC: lyso-phosphatidylcholine; PA: phosphatidic acid; PG: phosphatidylglycerol; PI: phosphatidylinositol; PS: phosphatidylserine; MGDG: monogalactosyldiacylglycerol; DGDG: digalactosyldiacylglycerol

Software

1. Analyst 1.5.1 (AB SCIEX)

Procedure

A. Total lipid extraction (Figure 1)

1. Aliquot 3 ml preheated isopropanol (75 °C with 0.01% BHT) into glass tubes (50 ml, 25 x 150 mm tubes with Teflon-lined screw caps are recommended).
2. Fresh *Arabidopsis* leaves were harvested and immediately soaked in hot isopropanol for at least 15 min in 75 °C water bath to inactivate phospholipase D. To achieve higher reliability and repeatability, 5-30 mg delipidated “dry weight” as measured in step A10 is recommended (~2-8 fully developed leaves to start with). Samples were cooled to room temperature after incubation. Isopropanol treated samples are stable at room temperature within a day.
3. Add 1.5 ml chloroform and 0.6 ml water to cooled samples, mix by vortexing. Incubate samples in water bath shaker at room temperature, 50 rpm for 1 h.
4. Transfer extracted lipids (disregard potential phase separation) with a glass Pasteur pipettes to another glass collection tube.
5. Add 4 ml lipid extraction solution to leaf samples and incubate in 25 °C water bath shaker for 30 min. Transfer extracts to the same collection tube.
6. Repeat step A5 to extract samples with lipid extraction solution until all leaf samples become white. It is recommended to extract each sample the same number of times. 5 extractions (including isopropanol in step A2) are usually needed. Old leaves are harder to be extracted completely, longer incubation time in steps A2, A3 and A5 and more extraction steps may help.
7. Add 1 ml KCl solution to the combined extract (~20 ml), mix thoroughly by vortex, centrifuge at ~200 x g for 10 min, discard upper phase carefully.
8. Add 2 ml water, mix by vortex, centrifuge at ~200 x g for 10 min, discard upper phase carefully.
9. Dry lipid samples completely in the nitrogen evaporator, mild heating (<50 °C) may be used to accelerate the process. Dried lipids can be stored at -80 °C with cap tightly closed. An aliquot of lipids can be resolved by thin layer chromatography (TLC) for quality check (Figure 2). Briefly, total lipids corresponding to ~0.2-1 mg DW were spotted on the TLC Silica gel 60 plate and resolved by chloroform:methanol:acetic acid:H₂O = 85:15:12.5:3.5. The plate was left dry completely in the fume hood before staining by iodine vapor and examining by eye or 0.01% primuline in acetone/H₂O (60:40; v/v) and examining the plates under ultraviolet (360 nm) light.
10. Dry extracted leaves in oven at 105 °C overnight. Weigh to get “dry weight”.

B. Lipid profiling by ESI-MS/MS and data processing

1. Reconstitute dried lipid samples (step A9) in chloroform, cap tightly before use to avoid evaporation. For easy sample preparation, lipid samples were diluted to the same concentration according to dry weight. We usually use 250 μ l chloroform per mg dry weight.
2. Each sample contains 840 μ l solution B, 5 μ l PL standard, 3 μ l GL standard and lipid sample corresponding to 0.1-0.2 mg dry weight (25-50 μ l), add chloroform to 1,200 μ l in a 2 ml snap seal autosampler vial, cap with pre-cut vial caps.
3. For every 10 lipid samples, only one "internal standard (IS)" vial is used for machine correction/contamination control. Chloroform is used in place of lipid samples in this vial.
4. Load the autosampler in the following format: IS1, sample1-10, IS2, sample11-20 and so on. "Blank" samples are recommended between different genotypes, which contains only solution B and chloroform.
5. Profile lipid samples by ESI-MS/MS. Detailed information can be found in Welti *et al.* (2002). The parameters we used on API 4000 are listed in section C. Optimization is needed on different setups, such that fewer or more parameters can be adjusted on older or newer machines. A representative phosphatidylethanolamine (PE) profiling result is attached (Figure 3).
6. Data were extracted by Analyst 1.5.1 with necessary corrections, including background subtraction and smooth. For each lipid class, multiple (usually 2) novel lipid species were used as internal standards (Table 1), linear interpolation according to molecular weight of each lipid species was used for internal standard peak area normalization. The amount of each lipid species was calculated according to sample peak area compared with normalized internal standard peak area. Five or more biological repeats are recommended, Student's t-test is used for statistical analysis.
7. Lipid data may be presented as absolute amount (nmol/mg "dry weight", recommended) or percentage of total lipids (PL and GL).

C. Lipid profiling parameters on API 4000

API 4000 LC-MS/MS system equipped with TurbolonSpray probe and HTC PAL autosampler was used for lipid profiling. The source temperature was 100 °C, interface heater was on, IonSpray Voltage was set to 5.5 kV, nitrogen gas was used for both source and curtain gas. The scan resolution was set to 0.1Da. The samples were introduced using the autosampler with 1 ml syringe and injection loop for full lipid profiling. The samples were presented to the TurbolonSpray probe by direct infusion at 0.03 ml/min. The scanning parameters for each lipid class were listed in Table 2.

Table 2. Scanning parameters

Lipid classes	Ion analyzed	Scan mode	Polarity	Time	Cycles	Declustering Potential	Entrance Potential	Collision energy	Exit Potential
PE/LYSOPE	[M+H] ⁺	Neutral Loss of 141 amu	+	3.1	37	100	15	28	11
PC/LYSOPC	[M+H] ⁺	Precursors of m/z 184 ⁺	+	1.7	10	120	14	40	14
PA	[M+NH ₄] ⁺	Neutral Loss of 115 amu	+	4	24	100	14	25	14
PG	[M+H] ⁺	Neutral Loss of 189 amu	+	3.5	21	100	14	20	14
PI	[M+H] ⁺	Neutral Loss of 277 amu	+	3	51	100	14	25	14
PS	[M+H] ⁺	Neutral Loss of 185 amu	+	3.5	42	100	14	25	14
MGDG	[M+H] ⁺	Neutral Loss of 179 amu	+	3.5	42	90	10	21	23
DGDG	[M+H] ⁺	Neutral Loss of 341 amu	+	3.5	42	90	10	24	23

PE: phosphatidylethanolamine; LYSOPE: lyso-phosphatidylethanolamine; PC: phosphatidylcholine; LYSOPC: lyso-phosphatidylcholine; PA: phosphatidic acid; PG: phosphatidylglycerol; PI: phosphatidylinositol; PS: phosphatidylserine; MGDG: monogalactosyldiacylglycerol; DGDG: digalactosyldiacylglycerol

Representative data

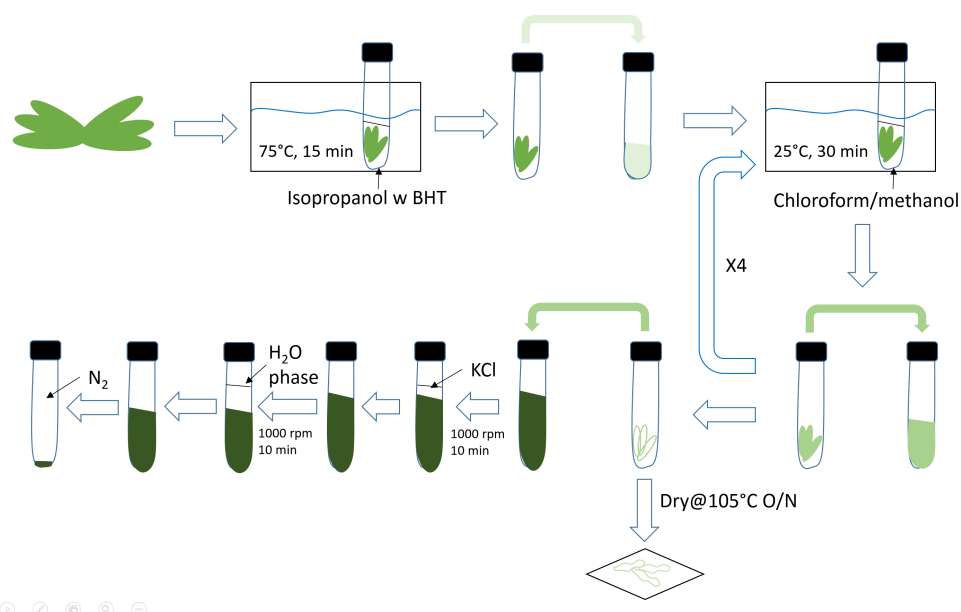


Figure 1. Illustration of lipid extraction workflow. 1,000 rpm (~200 x g).

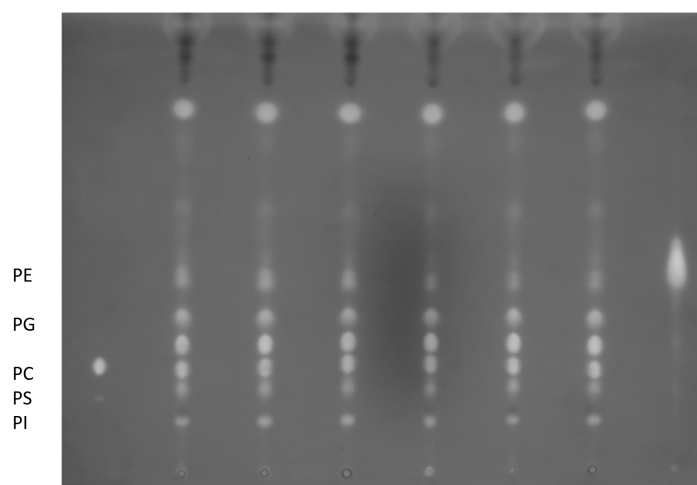


Figure 2. TLC separation of phospholipids. Lipids extracted in section A can be resolved on a TLC plate. The mobile phase used here is chloroform:methanol:acetic acid:H₂O = 85:15:12.5:3.5 (v/v).

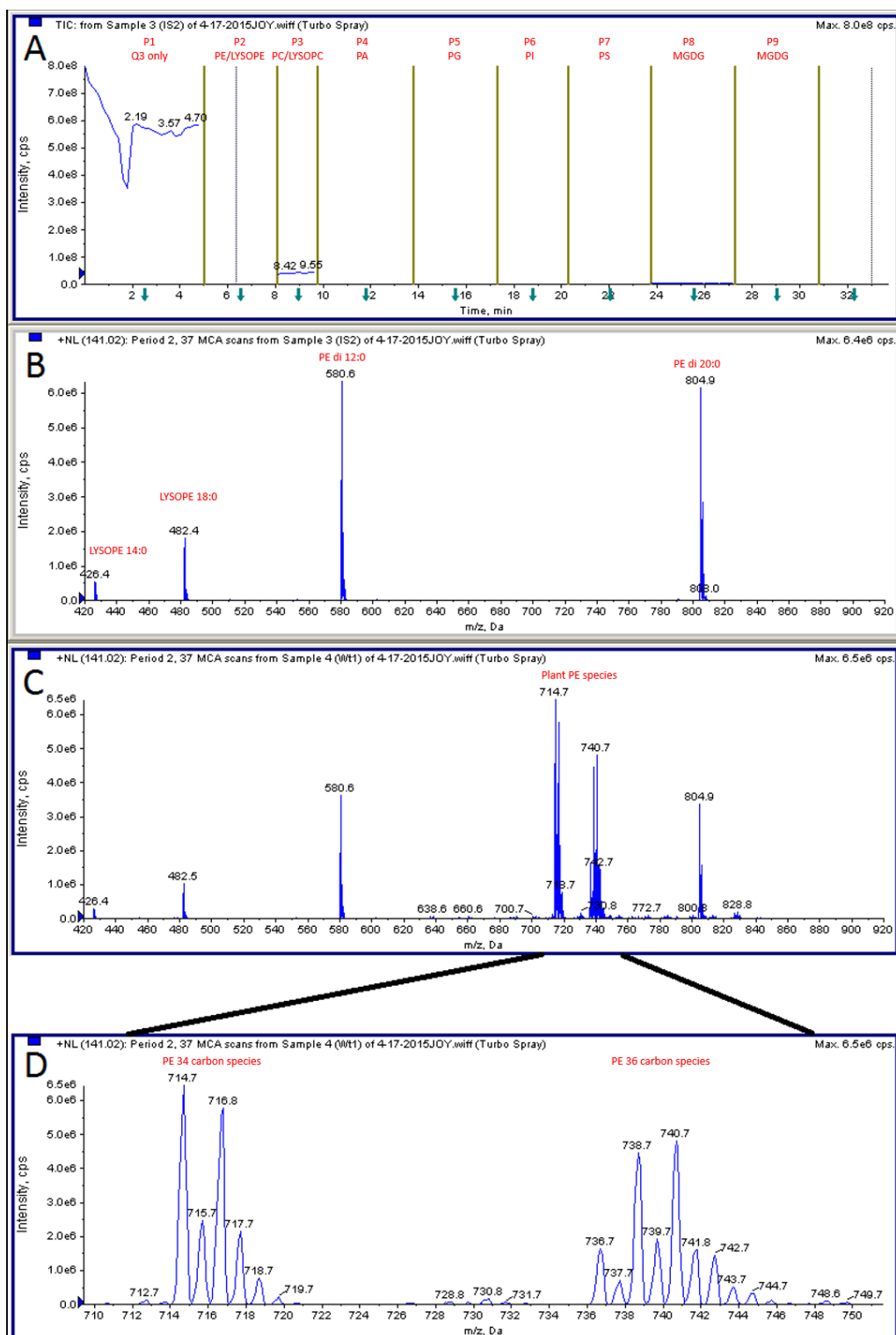


Figure 3. Representative MS data. A. Different lipid classes were profiled in tandem periods. Period 1 is for control purpose and not used in calculation. B. PE profile in IS sample. C. PE profile in plant sample. D. Enlarged view of plant PE species.

Notes

1. This Method can be easily adapted to roots, flowers, stems and whole siliques.
 - a. For *Arabidopsis* seed samples, the reproducibility of “dry weight” is highly dependent on seed stage and loss of samples during transferring lipid extracts. In this case, the percentage normalization is recommended.
 - b. For materials that are hard to extract completely, tissue disruption in liquid nitrogen before extraction is recommended. In this case, a centrifugation step (e.g., ~200 x g, 10 min) is needed each time before transferring lipid extracts.

Recipes

1. Isopropanol solution with BHT
 0.01% (w/v) BHT (~0.5 mM)
 Preheat at 75 °C
Note: stable at room temperature
2. Lipid extraction solution
 Chloroform:methanol = 2:1 (v/v), with 0.01% BHT
Note: stable if capped tightly at room temperature
3. KCl solution
 1 M KCl
Note: stable at room temperature
4. Solution B
 Methanol:NH₄Ac (300 mM) = 20:1(v/v)
Note: stable at room temperature
5. Galactolipid (GL) standard
Note: store at -80 °C, thaw and mix thoroughly before use
 Diluted in chloroform:methanol:H₂O = 35:65:8 [(v/v), or close to]
 MGDG (~0.5 nmol/μl)
 DGDG (~0.25 nmol/μl)
6. Phospholipid (PL) standard
Note: store at -80 °C, thaw and mix thoroughly before use
 Diluted in chloroform:methanol:H₂O = 35:65:8 [(v/v), or close to]
 See Table 1 for an example of PL standard content

Acknowledgments

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References

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