

Measuring the Interactions between Peroxisomes and Chloroplasts by *in situ* Laser Analysis

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[Abstract] Quantitative analysis has been necessary for deeply understanding characteristic of organelles function. This is the detailed protocol for the quantification of the physical interaction between peroxisomes and chloroplasts taken by laser scanning microscopy described by Oikawa *et al.* (2015). To clarify the morphological interactions between both organelles, we measured the contact length between two organelles (interaction length) in the fluorescent microscope image by using image analysis software ImageJ. The result clearly revealed that the contact length in light condition is much longer than that in dark condition. In addition, the force of the morphological interaction was quantified utilizing intersection technology of femtosecond laser and atomic force microscope (AFM). When an intense femtosecond laser is focused near the interface of two organelles, the adhesion is broken by a force due to the laser. The adhesion strength in light and dark conditions was estimated from the force calibrated by AFM. The detailed procedure is described in Bio-protocol as another protocol entitled “Quantification of the adhesion strength between peroxisomes and chloroplasts by femtosecond laser technology” (Hosokawa *et al.*, 2016). These methods can be applied to other physical interaction between different types of organelles such as nuclei, mitochondria, Golgi, and chloroplasts.

Materials and Reagents

1. Glass slide (super frost) (Matsunami Glass)

Note: Any types of glass slide suitable for fluorescence observation can be used. We attached black tape on the glass slide to envelop the sample with cover slip (see Figure 1D).

2. Cover slip (24 x 60 No.1, Thickness 0.12-0.17 mm) (Matsunami Glass)

3. 10 ml disposable syringe (Terumo Medical Corporation)
4. *Arabidopsis thaliana* (ecotype Columbia) expressing peroxisome-targeted GFP (GFP-PTS1) (Mano *et al.*, 2002)
Note: 1-4 is shown in Figure 1A.
5. 1/3 x Murashige and Skoog salts (MS) medium (Wako Pure Chemical Industries, catalog number: 392-00591) containing vitamins, 1% sucrose, and MES buffer (pH 5.7) (Wako Pure Chemical Industries, catalog number: 341-01622)
6. Agar powder for plant growth (Funakoshi, catalog number: BA-10)
7. Distilled water
8. KOH
9. Plant culture medium (see Recipes)

Equipment

1. Scissors and tweezers (shown in Figure 1A)
2. 40x dry system objective lens (ZEISS, EC Plan-Neofluar®)
3. Growth chamber for growing plants (100 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{sec}^{-1}$ white Light for 16 h and dark for 8 h, 22 °C)
4. Black box for dark condition
5. Confocal laser scanning microscope (ZEISS, model: LSM510 META)

Software

1. NIH ImageJ software 1.46 (<http://imagej.nih.gov/ij>)

Note: ImageJ is also available on Mac OS X, Windows and Linux.

Procedure

A. Preparation of leaf section

1. Cut the rosette leaves of 3-week-old *Arabidopsis* plants in 1 cm diameter and placed in a syringe filled with water (Figure 1B).
2. Plug the top of syringe by finger and pull plunger to deaerate the leaves (Figure 1B).
3. Mount one piece of the deaerated-leaf section (Figure 1C) on slide glass with water and cover with cover glass (Figure 1D).
4. Incubate the pretreated-leaf sample in the dark or light for 2 h in growth chamber.

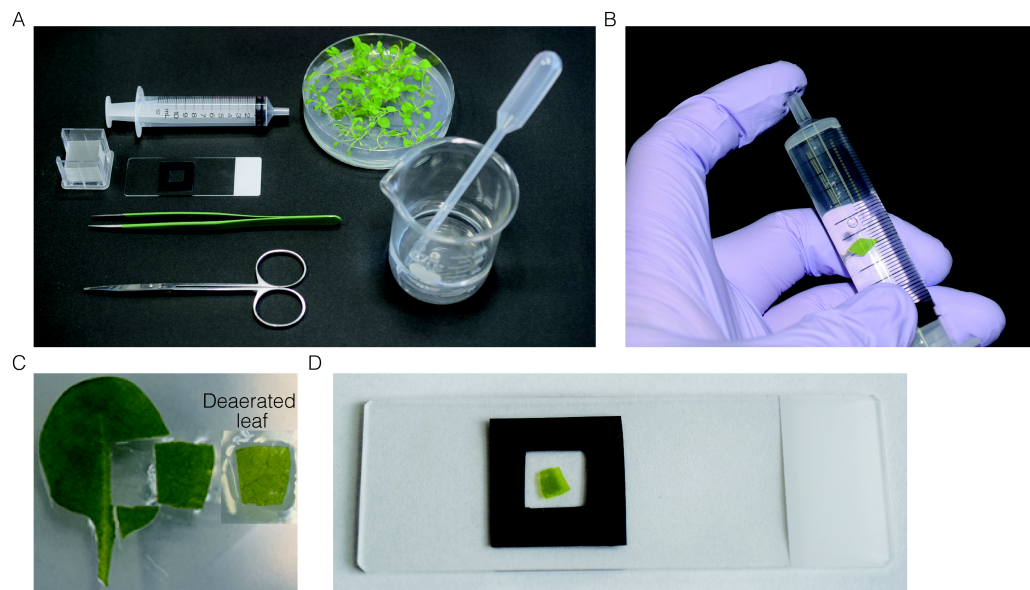


Figure 1. Preparation of leaf section for CLSM observation. A. All instruments used for preparation of leaf section. B. Deaeration of the leaves by pulling plunger while plugging the top of syringe. C. Leaf section used for observation. Before (left) and after (right) deaeration. D. Deaerated-leaf section mounted on the slide glass.

B. Taking organelles images by CLSM

1. The images of organelles in leaf palisade mesophyll cells are focused and obtained through a 40x dry system objective lens using LSM 510 (Figure 4).
2. To observe GFP fluorescence, a 488 nm Ar/Kr laser is used and the fluorescence is detected through an emission filter BP505-550. To detect the autofluorescence from chlorophyll, a 543 nm He/Ne laser is used and the signal is acquired through emission filters LP580.
3. Obtain both fluorescence simultaneously and save the data as a single image of tif. file.

Note: The observation of organelles should be finished within 30 min after moving the sample from growth chamber, since the change of light condition affects the organelle motility and interaction.

C. Quantification of the organelle interaction

1. Launch an Image J software and open the obtained images.
2. Set the scale of s: Analyze > Set Scale
3. Set the measurements: Analyze > Set Measurements > check the column of Shape descriptors (Figure 2).

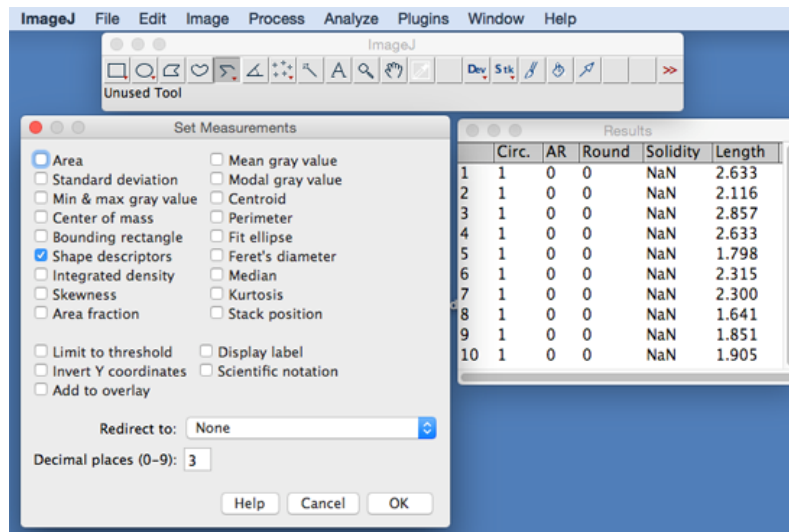


Figure 2. Analyze menu of ImageJ used to measure the length of segments lines. Choose Shape descriptors in Set Measurements window. Results of 10 interaction lengths are shown as an example of calculation after selecting Measure mode below Analyze menu.

- Using line tool, draw a line connecting two intersection points between two organelles, or using segment tool, draw the curve line (red line in the right panel of Figure 3 and lower panel of Figure 4) for measuring.

Note: The image is magnified by zoom (in) function of ImageJ when drawing a line. Image > Zoom > In{+}.

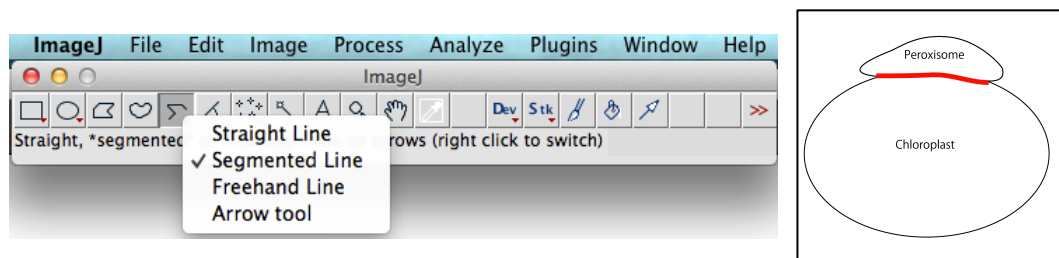


Figure 3. Segmented Line tool of ImageJ used to measure the interaction length (red line in the right panel) between a peroxisome and a chloroplast

- Measure the length of lines: Analyze > Measure

Note: An average of at least 50 interactions between peroxisomes and chloroplasts should be measured to calculate the average length in independent three experiments (Figure 4, Table 1). Here, we show results of ten samples for measurements of interaction lengths between peroxisomes and chloroplasts.

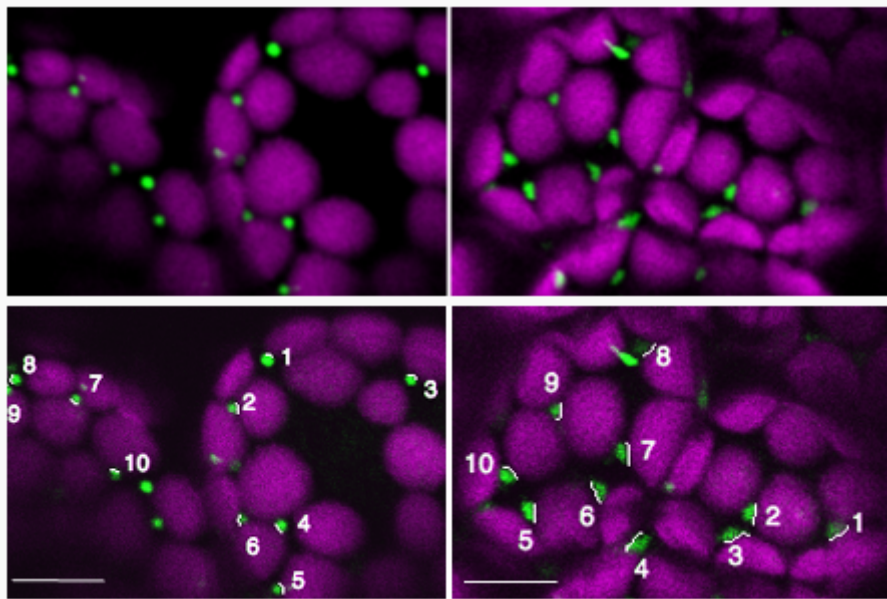


Figure 4. Measurements of the 10 selected-interaction lengths of membrane contact area between peroxisomes (green) and chloroplasts (magenta) in the dark (left panel) and light (right panel) at lower panels. Spherical peroxisomes in the dark change their shape to elongation in light, suggesting that their physiological interaction is enhanced in light. Upper panels are original images. Bars = 10 μ m

Table 1. Results of measurements of the 10 selected-contact lengths between peroxisomes and chloroplasts in dark and light. The contact length in light condition was longer than that in dark condition. This fact suggests that the peroxisomes assist photorespiration by tightly contacting with the chloroplasts in photosynthesis.

	Dark (μ m)	Light (μ m)
1	0.78	2.63
2	1.23	2.12
3	1.23	2.86
4	1.15	2.63
5	1.33	1.80
6	1.23	2.32
7	0.87	2.30
8	1.06	1.64
9	1.51	1.85
10	0.96	1.91
Average	1.13	2.20
Stdev	0.22	0.41

Recipes

1. Plant culture medium

Dissolve 10 g sucrose, 1.53 g MS salt, and 0.5 g MES in 1 L pure water, and then adjust the pH to 5.7 with 1 M KOH

Add 8 g agar in the medium, and then autoclave

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