

Telomerase Repeated Amplification Protocol (TRAP)

Ilgen Mender¹ and Jerry W. Shay^{1, 2*}

¹Department of Cell Biology, University of Texas Southwestern Medical Center, Dallas, USA;

²Center for Excellence in Genomics Medicine Research, King Abdulaziz University, Jeddah, Saudi Arabia

*For correspondence: Jerry.Shay@utsouthwestern.edu

[Abstract] Telomeres are found at the end of eukaryotic linear chromosomes, and proteins that bind to telomeres protect DNA from being recognized as double-strand breaks thus preventing end-to-end fusions (Griffith *et al.*, 1999). However, due to the end replication problem and other factors such as oxidative damage, the limited life span of cultured cells (Hayflick limit) results in progressive shortening of these protective structures (Hayflick and Moorhead, 1961; Olovnikov, 1973). The ribonucleoprotein enzyme complex telomerase-consisting of a protein catalytic component *hTERT* and a functional RNA component *hTR* or *hTERC*- counteracts telomere shortening by adding telomeric repeats to the end of chromosomes in ~90% of primary human tumors and in some transiently proliferating stem-like cells (Shay and Wright, 1996; Shay and Wright, 2001). This results in continuous proliferation of cells which is a hallmark of cancer. Therefore, telomere biology has a central role in aging, cancer progression/metastasis as well as targeted cancer therapies. There are commonly used methods in telomere biology such as [Telomere Restriction Fragment \(TRF\)](#) (Mender and Shay, 2015b), Telomere Repeat Amplification Protocol (TRAP) and [Telomere dysfunction Induced Foci \(TIF\) analysis](#) (Mender and Shay, 2015a). In this detailed protocol we describe Telomere Repeat Amplification Protocol (TRAP).

The TRAP assay is a popular method to determine telomerase activity in mammalian cells and tissue samples (Kim *et al.*, 1994). The TRAP assay includes three steps: extension, amplification, and detection of telomerase products. In the extension step, telomeric repeats are added to the telomerase substrate (which is actually a non-telomeric oligonucleotide, TS) by telomerase. In the amplification step, the extension products are amplified by the polymerase chain reaction (PCR) using specific primers (TS upstream primer and ACX downstream primer) and in the detection step, the presence or absence of telomerase is analyzed by electrophoresis. TSNT is, an internal standard control, amplified by TS primer. NT is its own reverse primer, which is not a substrate for telomerase. These primers are used to identify false-negative results by if the gel lacks internal control bands.

Materials and Reagents

1. Cancer cells (H1299 non-small cell lung, A549 non-small cell lung cancer cells)
2. Tris-HCl (pH 8.3 and pH 8.0)

3. Magnesium Chloride (MgCl₂) (Thermo Fisher Scientific, catalog number: BP241)
4. Potassium chloride (KCl) (Sigma-Aldrich, catalog number: P9541)
5. Tween™ 20 (Thermo Fisher Scientific, catalog number: BP337)
6. Ethylene glycol-bis(2-aminoethylether)-N, N, N', N'-tetraacetic acid EGTA (Sigma-Aldrich, catalog number: E3889)
7. Ethylenediaminetetraacetic Acid (EDTA) (Thermo Fisher Scientific, catalog number: BP-120)
8. Nonidet-P40 (Fluka BioChemika, catalog number: 74385)
Note: Currently, it is "Sigma-Aldrich, catalog number: 74385".
9. Glycerol (Sigma-Aldrich, catalog number: G5516)
10. 2-mercaptoethanol (Sigma-Aldrich, catalog number: M3148)
11. 4-(2-Aminoethyl) benzenesulfonyl fluoride hydrochloride (AEBSF) (Sigma-Aldrich, catalog number: A8456)
12. dNTP (Invitrogen)
13. Cy5-TS primer (Sigma-Aldrich)
14. ACX (reverse primer) (Sigma-Aldrich)
15. TSNT (36-bp internal standard control) (Integrated DNA Technologies)
16. NT (reverse primer for internal standard) (Sigma-Aldrich)
17. Takara Taq DNA polymerase, hot start version (Takara Bio Company, ClonTech, catalog number: R007A)
18. 40% acrylamide and bis-acrylamide solution (19:1) (Bio-Rad Laboratories, catalog number: 161-0144)
19. Ammonium persulfate (APS) (Bio-Rad Laboratories, catalog number: 161-0700)
20. N, N, N', N'-Tetramethylethylenediamine (TEMED) (Sigma-Aldrich, catalog number: T9281)
21. Primer mix (see Recipes)
22. Cy5-TS primer (see Recipes)
23. 10x TRAP reaction buffer (see Recipes)
24. 50x dNTP (see Recipes)
25. NP-40 lysis buffer (see Recipes)

Equipment

1. Polymerase Chain Reaction (PCR) Thermo Cyclor (Bio-Rad Laboratories, model: PTC-1148)
2. Typhoon PhosphorImager® scanner system (Amersham Biosciences, GE Healthcare, model: Typhoon TRIO)

Software

1. ImageQuant Software (Molecular Dynamics)

Procedure

A. Prepare cell lysates

1. Collect 100,000 cells into a DNase/RNase -free microfuge tube.
2. Centrifuge cells at 3,000 x g for 5 min.
3. Remove the supernatant.

Note: The pellet doesn't need to be washed. If samples are to be used at a later time point, put in -80 °C.

4. Resuspend cell pellet on ice-cold NP-40 lysis buffer at a concentration of 2,500 cells per μ l (40 μ l lysis buffer for 100,000 cells).
5. Leave on ice for 30 min or snap-freeze cell lysates in liquid nitrogen and then place at -80 °C.

B. Preparation for PCR-Cy5 fluorescent gel-based TRAP

1. Keep samples on ice.
2. Choose a positive control (*i.e.* HCT116, H1299) and prepare three different serial dilutions (1:10) in NP-40.
3. Prepare the TRAP master mix. Final volume of the PCR reaction is 50 μ l.

Master mix for one sample:

H ₂ O	40.2 μ l
Trap buffer	5 μ l
dNTP	1 μ l
Cy5-TS primer	1 μ l
Primer mix	1 μ l
BSA	0.4 μ l
Taq polymerase	0.4 μ l

4. Add 1 μ l of sample to 49 μ l of the master mix.

Note: For negative control, 1 μ l NP-40 to 49 μ l of the master mix.

C. Amplification of the extension products by PCR

1. 25 °C for 40 min for extension.
2. 95 °C for 5 min to deactivate telomerase.
3. 24 to 29 cycles at:
 - 95 °C for 30 sec
 - 52 °C for 30 sec
 - 72 °C for 45 sec

4. 72 °C for 10 min

5. 4 °C for up to five days

Note: Samples can be stored at -20 °C for 1 month.

D. Running acrylamide gel

1. Add 5 µl of loading dye to each sample.

2. Making the 10% nondenaturing acrylamide gel.

32.5 ml H₂O (MilliQ® water)

12.5 ml 40% acrylamide (19:1 acrylamide:bisacrylamide)

5 ml 5x TBE

250 µl 10% APS

50 µl TEMED

3. 0.5x TBE running buffer

4. Running time is about 2.5 h at 200/220 volts.

5. Load 25 µl of the sample to each well.

6. Visualize using Typhoon® that can read Cy5 fluorescein.

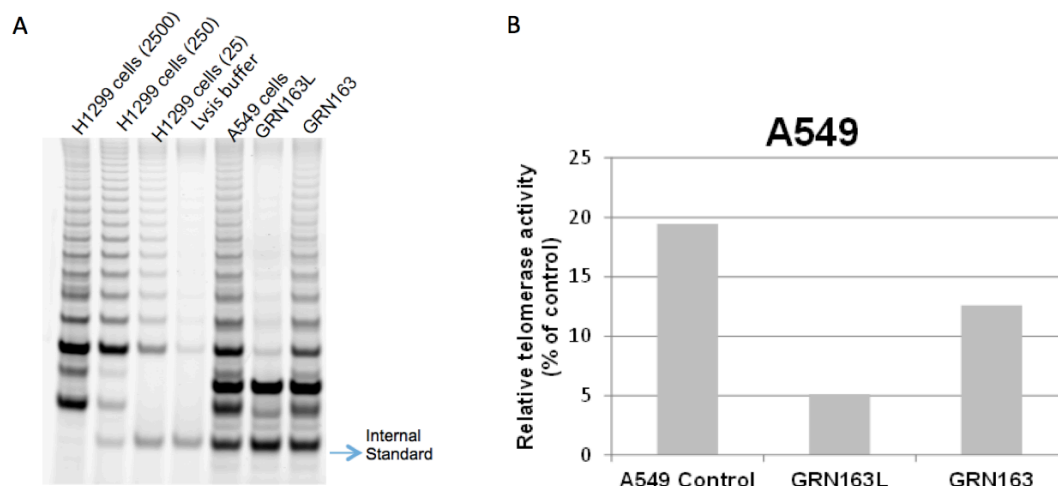


Figure 1. TRAP gels show partial telomerase inhibition with 10 µM GRN163L in A549 non small cell lung cancer cells for 72 h. It is known that a lipid modified N3→P5 thio-phosphoramidate oligonucleotide, GRN163L (also referred to as Imetelstat), inhibits telomerase more potently than its parental non-conjugated thio-phosphoramidate sequence, GRN163, in cancer cells (Herbert *et al.*, 2005). This figure also shows that GRN163L (L= lipidated with palmitoyl C16) inhibits A549 cells more potently compared to GRN163. H1299 non-small lung cancer cells were used with three different dilutions as a positive control. 2500, 250 and 25 cells for H1299 were used in lane 1, lane 2 and lane 3, respectively. No cells were added in lysis buffer in lane 4, which is a negative control (A). ImageQuant Software was used to determine the intensity of the telomerase products (6 bp-ladder) and the ITAS (Internal Standard Control, 36 bp) band. The ratio of intensity

- c. Clean the outside of the tubes and rack with diluted bleach (spray 10% bleach on the tubes, then dry them with paper towel or spray 10% bleach on paper towel and clean the outside of the tubes).
- d. Return to the TRAP area with the tube and prepare aliquots to store at -20 °C up to 1 year.

2. Cy5-TS primer

TS oligo is purchased modified with Cy5 on the 5' end (HPLC or PAGE purified). Final concentration of TS primer is 100 ng/μl (diluted in DEPC water).

Sequence: 5'-AAT CCG TCG AGC AGA GTT-3'

3. 10x TRAP reaction buffer

	Final concentration
Tris-HCl (pH 8.3)	200 mM
MgCl ₂	15 mM
KCl	630 mM
Tween 20	0.5% (v/v)
EGTA	10 mM

4. 50x dNTP

Use a mix containing all four DNA nucleotides (dTTP, dATP, dCTP, dGTP) at an equivalent and final concentration of 2.5 mM

5. NP-40 lysis buffer

	Final concentration
Tris-HCl (pH 8.0)	10 mM
MgCl ₂	1 mM
EDTA	1 mM
NP-40	1% (v/v)
Sodium deoxycholate	0.25 mM
Glycerol	10% (v/v)
NaCl	150 mM
2-mercaptoethanol	5 mM
AEBSF	0.1 mM

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