

XTT Assay of Antifungal Activity

Flávio V. Loures^{1*} and Stuart M. Levitz²

¹Department of Medicine, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America, Department of Immunology, Institute of Biomedical Sciences, São Paulo University, São Paulo, Brazil; ²Department of Medicine, University of Massachusetts Medical School, Worcester, Massachusetts, USA

*For correspondence: loures@icb.usp.br

[Abstract] XTT assay is a colorimetric method that uses the tetrazolium dye, 2,3-bis-(2-methoxy-4-nitro-5-sulphenyl)-(2H)-tetrazolium-5-carboxanilide (XTT) to quantify cell-mediated damage to fungi. Actively respiring fungal cells convert the water-soluble XTT to a water-soluble, orange colored formazan product (Meshulam *et al.*, 1995). Here, we describe the protocol that measures the ability of plasmacytoid dendritic cells (pDCs) to exert antifungal activity. This approach was first established with human polymorphonuclear cells (PMN) by Meshulam *et al.* (1995) and then adapted to pDC by Ramirez-Ortiz *et al.* (2011) and Loures *et al.* (2015). It can be modified for use with other effector cells and to test compounds for antifungal activity.

Materials and Reagents

1. *Aspergillus fumigatus* (*A. fumigatus*) clinical isolate Af293 (Nierman *et al.*, 2005)
2. Human pDCs obtained from healthy donors
3. RPMI-1640 medium without phenol red (Life Technologies, Gibco®, catalog number: 11875119)
4. Penicillin G/ streptomycin sulfate (Life Technologies, Gibco®, catalog number: 15140122)
5. L-glutamine (Life Technologies, Gibco®, catalog number: 25030081)
6. Sodium pyruvate (Life Technologies, Gibco®, catalog number: 11360070)
7. Sterile distilled water (Life Technologies, Gibco®, catalog number: 10977015)
8. Sterile PBS (Corning Incorporated, catalog number: 21040CM)
9. 96 well half area plates (Corning, Costar®, catalog number: 07200309)
10. Tips
11. 15 ml Centrifuge tube (Falcon®, catalog number: 352059)
12. XTT (Sigma-Aldrich, catalog number: X4251)
13. Coenzyme Q (Sigma-Aldrich, catalog number: D9150)
14. Tween 20 (Thermo Fisher Scientific, catalog number: BP337500)
15. Sabouraud dextrose agar (Thermo Fisher Scientific, catalog number: R454472)
16. Supplemented media (see Recipes)

17. Slants (see Recipes)

Equipment

1. 30 μ m Nylon mesh (Genesee Scientific Corporation, catalog number: 57105)
2. 10 ml Syringe (BD Bioscience, catalog number: 305482)
3. Inverted microscope (Nikon)
4. Pipettes
5. Multichannel pipette
6. *Neubauer* chamber
7. Reagents reservoir (Corning, Costar®, catalog number: 07200127)
8. Microplate reader with capacity to measure OD₄₅₀ and OD₆₅₀ (Versamax)
9. Water bath (Thermo Fisher Scientific)
10. CO₂ Incubator (37 °C)
11. Vortex (Fisher Vortex Genie 2)(Scientisfic Industries, Inc., catalog number: 12812)
12. Autoclavec

Procedure

1. Grow *A. fumigatus* (Af293) on Sabouraud dextrose agar slants for 3-5 days at 37 °C. The culture will turn white as hyphae grow and then green as sporulation (conidia formation) occurs.

Note: The assay can be adapted for other species of fungi.

2. Harvest the conidia with 2-3 ml of PBS containing 0.05% Tween 20 (PBS T20) gently removing the conidia from the slants with a tip. The conidia can be kept at 4 °C for up to 7 days before use in assays. Maintain sterile conditions when working with conidia.
3. Suspend the conidia by mixing on a vortex for few seconds. Draw the suspension into a 10 cc syringe. Fold 30 μ m nylon mesh inside the top of a 15 ml centrifuge tube. Filter the conidial suspension drop wise through the nylon mesh. Remove and discard the nylon mesh from the centrifuge tube.
4. Wash the conidia three times by suspension in 10 ml PBS T20 followed by centrifugation at 3,000 rpm/10 min.
5. Suspend the filtered conidia in 5 ml supplemented media.
6. Count the conidia in a *Neubauer* chamber.
7. Add 5×10^3 *A. fumigatus* conidia in 96-well half area plates in 100 μ l supplemented media per well. If desired for a dose response curve, the number of conidia per well can be varied; however, fewer conidia may not give adequate OD readings.
8. Keep the plates in the dark at room temperature (21 °C) in static conditions for 16 h in supplemented media to swell the conidia and then incubate for an additional 3 h at 37 °C to promote germination of hyphae. Note that after putting the plates in the incubator

check the germination process every 15 min until you get hyphae of 10-20 µm average length.

9. Withdraw the plate from the incubator when you get hyphae of 10-20 µm average length. If your pDCs are not ready yet keep the plate at 4 °C to avoid overgrowing of the hyphae. We suggest 2 h as the maximum period of storage at 4 °C.
10. Gently wash the hyphae 2 times with sterile PBS (200 µl).
11. Add 5×10^4 pDCs to the hyphae in a final volume of 100 µl supplemented media. Note that you can use a different number of pDCs or another effector cell type. Generally, each condition is tested in at least triplicate. Keep the co-culture under sterile conditions.
12. To determine background activity, 4 to 5 wells should contain pDC only.
13. To determine XTT reduction by fungi in the absence of pDC, 4 to 5 wells should be left with *A. fumigatus* hyphae only.
14. To determine blank values, 4 to 5 wells should have media only.
15. Following 2 h incubation at 37 °C, the pDCs are subjected to hypotonic lysis by three washes with 100 µl distilled water. It is critical that the washes are performed very gently and carefully to avoid removing hyphae.
16. Incubate for 30 min with 100 µl distilled water at 37 °C.
17. During the 30 min of incubation, the RPMI-1640 containing 400 µg/ml of XTT and 50 µg/ml of Coenzyme Q must be prepared. Stock solutions each containing 10 mg/ml XTT and Coenzyme Q should be prepared individually in PBS. Both compounds need high temperature to dissolve. Usually a 1 min incubation in a 60 °C water bath is sufficient. When more than 1 min is necessary, it is important to take the solutions out of the water bath (at 60 °C) immediately after they dissolve. The XTT and Coenzyme Q should be added to the RPMI-1640 a few minutes before adding to the wells. As phenol red can interfere with colorimetric readings, the RPMI-1640 should not contain phenol red.
18. Remove the supernatants, again taking great care taken not to remove the hyphae.
19. 100 µl RPMI media containing 400 µg/ml of XTT and 50 µg/ml of Coenzyme Q, are added, and the wells are incubated for 2 h at 37 °C.
20. The OD₄₅₀ and OD₆₅₀ are measured on the microplate reader.
21. Data are expressed as percent antifungal activity according to the formula:

$$\% \text{Antifungal Activity} = \left[1 - \left(\frac{OD_{Af+pDCs} - OD_{pDCs}}{OD_{Af} - OD_{Blank}} \right) \right] \times 100$$

OD_{Af + pDC} is (OD₄₅₀ - OD₆₅₀) of wells containing *A. fumigatus* hyphae with pDCs.

OD_{pDC} is (OD₄₅₀ - OD₆₅₀) of wells containing pDCs.

OD_{Af} is (OD₄₅₀ - OD₆₅₀) of wells containing *A. fumigatus* hyphae alone.

OD_{blank} is (OD₄₅₀ - OD₆₅₀) of wells containing media alone.

Recipes

1. Supplemented media
RPMI-1640 (without phenol red) with 100 U/ml penicillin, 100 U/ml streptomycin, 2 mM L-glutamine, and 1 mM sodium pyruvate
2. Slants
Add 4.5 of Sabouraud dextrose agar to 100 ml distilled water
Autoclave it for 15 min
Add 3 ml per 15 ml tube
Tilt the tube at a 45° to 60° angle and let it solidify. Keep at 4 °C until use.

Acknowledgments

This work was supported in part by National Institutes of Health Grants RO1HL112671, RO1AI025780, and RO1AI072195. FVL acknowledges support from Fundação de Amparo Pesquisa de São Paulo (FAPESP) and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq).

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