

Ex vivo Human Natural Killer (NK) Cell Stimulation and Intracellular IFN γ and CD107a Cytokine Staining

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[Abstract] Natural killer (NK) cells comprise 5–20% of peripheral blood mononuclear cells (PBMC) in humans. In addition to their fundamental roles in the defense against viral infections and tumor surveillance, NK cells help shape adaptive immune responses through their production of cytokines. NK cells are traditionally identified as CD3^{neg}, CD14^{neg}, CD19^{neg} lymphocytes expressing CD56. Using a combination of markers that includes CD56 and CD7 greatly increases the ability to define the phenotype and function of NK cell subsets. Two key markers of NK cell function are the production of IFN γ and the release of cytotoxic granules measured by the expression of CD107a. Here we describe a method to assess IFN γ and CD107a expression in NK cells following stimulation with target cells or cytokines. This method can be used to assess the general functional capacity of NK cells in peripheral blood mononuclear cells from a wide range of study participants.

Materials and Reagents

1. Alexa700-conjugated mouse anti-human CD7 clone 124-1D1 (eBioscience, catalog number: 56-0079-42)
2. K562 cell line (Kindly provided by Dr. Lewis L. Lanier, University of California, San Francisco, USA)
Note: These can also be purchased from ATCC, catalog number CCL-243.
3. Phycoerythrin (PE)-Texas Red (ECD)-conjugated mouse anti-human CD3 clone UCHT1 (Beckman Coulter, catalog number: IM2705U)
4. ECD-conjugated mouse anti-human CD14 clone RMO52 (Beckman Coulter, catalog number: IM2707U)
5. PE-Cy7-conjugated mouse anti-human CD56 clone NCAM16.2 (BD Biosciences, catalog number: 335791)
6. Pacific Blue-conjugated mouse anti-human CD16 clone 3G8 (BD Biosciences, catalog number: 558122)
7. APC-Cy7-conjugated mouse anti-human CD19 clone SJ25C1 (BD Biosciences, catalog number: 557791)
8. Fluorescein isothiocyanate (FITC)-conjugated mouse anti-human CD107a clone H4A3 (BD Biosciences, catalog number: 555800)

9. APC-conjugated mouse anti-human IFN γ clone B27 (BD Biosciences, catalog number: 554702)
10. Human IgG (Sigma-Aldrich, catalog number: I4506)
11. Anti-mouse immunoglobulin G-coated compensation beads (BD Biosciences, catalog number: 552843)
12. Amine Aqua Reactive Dye (AARD) (Life Technologies, catalog number: L34957)
13. 96 well U bottom plate (Corning, catalog number: 353077)
14. 96 well V bottom plate (Corning, catalog number: 3894)
15. RPMI (Life technologies, catalog number: 11875)
16. L-Glutamine 200 mM (100x) (Life technologies, catalog number: 25030)
17. Penicillin (10,000 Units/ml)-Streptomycin (10,000 μ g/ml) (Life technologies, catalog number: 15140)
18. Fetal bovine serum (Hyclone, catalog number: SH30071)
19. Buffy coats from Stanford Blood Center used to obtain Peripheral Blood Mononuclear Cells (PBMC)
20. Ficoll-Paque Premium (GE Healthcare, catalog number: 17-5442-03)
21. Cryopreserved PBMC samples from San Francisco based HIV-1 infected cohorts SCOPE and OPTIONS
22. Recombinant IL-12 (Peprotech, catalog number: 200-12)
23. Recombinant IL-18 (MBL & Biological Laboratories, catalog number: B001-5)
24. Brefeldin A from *Penicillium brefeldianum* (Sigma-Aldrich, catalog number: B7651)
25. BD golgi stop protein transport inhibitor containing monensin (BD Biosciences, catalog number: 554724)
26. Phosphate buffered saline (PBS) (Corning, catalog number: 21-040-CV)
27. Ethylenediaminetetraacetic Acid (EDTA) (Teknova, catalog number: E0306)
28. Bovine Serum Albumin (BSA) (Gemini BioProducts, catalog number: 700-100P)
29. 16% Paraformaldehyde (PFA) (Electron Microscopy Science, catalog number: 15710)
30. BD FACS Permeabilizing solution 2 (BD Biosciences, catalog number: 340973)
31. Cell Culture Grade Water (HyClone, catalog number: SH30529.02)
32. Deoxyribonuclease (DNase) I (Sigma-Aldrich, catalog number: DN25)
33. 15 ml conicals (Thermo Fisher Scientific, catalog number: 05-539-5)
34. Trypan blue in PBS (0.4% w/v) (Corning, catalog number: 25-900-CI)
35. Complete media (see Recipes)
36. FACS buffer (see Recipes)
37. Paraformaldehyde recipe (see Recipes)

Equipment

1. Biosafety cabinet (Nuaire, model: 407FM600)
2. 37 °C water bath (Cole Parmer)
3. Calibrated single-channel and multi-channel pipettes
4. Pipet-aid
5. Tips (10 μ l, 20 μ l, 200 μ l, 1,000 μ l)
6. Centrifuge (Beckman Coulter, Allegra 6R, rotor GH-3.8)
7. 37° Celsius Incubator (Thermo Forma, model: 3110)
8. Nikon Optiphot microscope for counting cells
9. Hemacytometer for counting cells (Hausser Scientific, catalog number: 1490)
10. 4-laser (405 nm, 488 nm, 532 nm and 633 nm) BD LSR-II

Software

1. FlowJo Single Cell Analysis software

Procedure

1. Recovery of Peripheral Blood Mononuclear Cells (PBMCs) from cryostorage.
 - a. Warm complete media in a 37 °C water bath prior to thawing.
 - b. Transfer frozen vials of cryopreserved PBMC from a cryofreezer into a 37 °C water bath and gently move the tube back and forth in the water, allowing the contents of the vial to thaw until there is only a small amount left frozen. Do not leave the cryovial unattended during the thawing process. Thawing will take approximately 1-2 min.
 - c. In a biosafety cabinet, remove the liquid from the cryovial and place into a 15 ml conical tube using a 2 ml serological pipette and pipettor.
 - d. Slowly add 10 ml of pre-warmed (37 °C) media dropwise into the 15 ml conical tube. Gently flick the tube as you are adding media to ensure that the liquid is well mixed.
 - e. Cap the tube and centrifuge at 330 x g for 5 min in a pre-cooled 4 °C centrifuge.
 - f. Decant supernatant, break up the pellet by flicking the tube, and resuspend in 10 ml of complete media containing DNase (0.01 mg/ml) to count cells.
 - g. Mix 10 μ l of cells with 10 μ l of Trypan blue. Take 10 μ l of that mixture and place onto a hemacytometer for counting.
 - h. Centrifuge cells at 330 x g for 5 min at 4 °C, decant supernatant, break up the cell pellet by flicking the tube, and resuspend in complete media containing DNase at a concentration of 5 million cells per ml.

2. Plate 500,000 PBMC per well in 100 μ l by single channel pipette in a 96 well U bottom plate.

Note: Alternative protocol for stimulation: Thawed PBMC can be incubated overnight (16-20 h) at 37 °C and 5% CO₂ in complete media supplemented with 200 IU/ml human recombinant IL-2 (NCI BRB Preclinical Repository). This significantly increases the responsiveness of the NK cells to stimulation with target cells; however, culturing overnight in IL-2 can also alter the phenotype of the NK cells. One needs to consider what the desired readout for the experiment is in choosing between the main protocol and this alternative approach.

3. Add FITC-conjugated anti-CD107a antibody using a single channel pipette at a dilution of 1:50 (*i.e.* 4 μ l in 200 μ l culture conditions) to all wells of a 96 well plate containing cells.

Notes:

- a. *The dilution of this antibody should be titrated for use with a particular cytometer.*
- b. *The final volume in each well should be 200 μ l. There will be 100 μ l of PBMC and either 100 μ l of target cells (K562) or 100 μ l of cytokine mixture.*
4. For target cell stimulation (*i.e.* K562), count and suspend 500,000 K562 cells in 1 ml of complete media containing DNase and add 50,000 target cells (100 μ l) using a single channel pipette to appropriate wells.
5. For cytokine stimulation, add IL-12 and IL-18 at a final concentration of 10 ng per cytokine per well. Add 1 μ l of each cytokine to 998 μ l complete media containing DNase. Add 100 μ l per well using a single channel pipette to appropriate wells for stimulation.
6. Mix all wells with a multichannel pipette by carefully pipetting up and down 3 times being careful not to cause bubbles.
7. Place plate in incubator for 1 h at 37 °C and 5% CO₂.
8. After 1 h, centrifuge plate at 330 x g in a 25 °C centrifuge.
9. Remove 50 μ l supernatant from each well being careful not to disturb cell pellet. To not disturb the pellet, tilt the plate at a 45-60° angle, and using a multichannel pipette, place tip to the side of the well and remove liquid slowly. The supernatant can be discarded.
10. Prepare a master mix containing Brefeldin A (BFA) and GolgiStop (monensin) mixture.
 - a. Dilute GolgiStop 7.8 μ l into 42.2 μ l complete media containing 0.01mg/ml DNase.
 - b. Make a master mix of: 20 μ l 10 mg/ml Brefeldin A (BFA)
20 μ l of diluted GolgiStop mixture from step 10a.
4,960 μ l complete media containing DNase
 - c. Plate 50 μ l master mix into each well using a single channel pipette. Mix well using a multichannel pipette.
11. Incubate another 5 h at 37 °C, 5% CO₂.

12. Transfer cells to a 96 well V bottom plate. Centrifuge plate at 330 x g for 5 min at 4 °C and remove supernatant. Flick supernatant into a container to be discarded. To flick, turn the plate over and give one firm shake.
13. Resuspend cells in 200 μ l FACS buffer using a multichannel pipette and centrifuge plate at 330 x g for 5 min at 4 °C.
14. Wash plate again adding 200 μ l FACS buffer per well using a multichannel pipette. Centrifuge plate at 330 x g for 5 min at 4 °C. Flick supernatant into a container to be discarded. To stop the protocol overnight at this point, see step 15. To continue, skip to step 16.
15. For time considerations, the protocol can now be stopped here overnight. Resuspend cells in 200 μ l FACS buffer using a multichannel pipette, place in the refrigerator overnight, and stain the next day. Prior to staining on Day 2, centrifuge plate at 330 x g for 5 min at 4 °C. Flick out supernatant. Blot the plate on paper towels. Then continue with step 16.
16. Surface stain cells.
 - a. Make up a master mixture of surface antibodies diluted in FACS buffer that have previously been titrated for your flow cytometer. Staining is performed in 50 μ l volume for 500,000 cells.
 - i. We have found the following combination works well for our BD LSR-II.

Antigen	Fluorophore	Clone	Dilutions
CD3	ECD	UCHT1	1:50
CD14	ECD	RMO52	1:400
CD19	APCCy7	SJ25C1	1:100
CD7	Alexa700	124-1D1	1:50
CD16	Pacific Blue	3G8	1:100
CD56	PECy7	NCAM16.2	1:100
IgG (100 μ g/mL)			1:10
Live/Dead Marker	Amine Aqua Reactive Dye (AARD)		1:200
FACS buffer			

- ii. The use of fluorescence minus one (FMOs) is recommended. These can be made using the exact same mixture of antibodies but omitting one antibody. We focus on FMOs for CD107a and IFN γ , although FMOs can be used for all fluorophores initially to set appropriate gating of cytometry data.
- iii. Make sure to have centrifuged the plate at 330 x g for 5 min at 4 °C and flicked out the supernatant prior to adding the surface stain. Aliquot 50 μ l of antibody cocktail into each well using a single channel pipette. Mix well by pipetting up

and down using a multichannel pipette. Allow plate to incubate on ice for 30 min to stain.

17. After 30 min, add 150 μ l FACS buffer to all wells using a multichannel pipette, centrifuge plate at 515 x *g* for 5 min at 4 °C. To improve recovery of cells from this step forward, a faster centrifuge speed is used.
18. Flick supernatant into a container to be discarded and blot the plate on paper towels.
19. Fix cells by adding 100 μ l per well of 2% paraformaldehyde in PBS on ice for 20 min using a multichannel pipette.
20. Spin at 915 x *g* in a 4 °C centrifuge for 5 min.
21. Flick supernatant into a container to be discarded and blot on paper towels.
22. Permeabilize cells using BD FACS Permeabilizing solution 2.
 - a. Perm2 is supplied as a 10x solution. Dilute to 1x using cell culture grade water.
 - b. Add 75 μ l of the 1x Perm2 solution per well using a multichannel pipette and pipet up and down to resuspend cell pellet.
 - c. Incubate plate for 10 min only at room temperature.
 - i. Shorter incubations will reduce intracellular staining while incubating longer than 10 min will significantly affect cellular integrity and result in loss of sample.
23. Wash once by adding 125 μ l FACS buffer to each well using a multichannel pipette.
24. Centrifuge plate at 915 x *g* in a 4 °C centrifuge for 5 min.
25. Flick supernatant into a container to be discarded and blot the plate on paper towels.
26. Intracellularly stain cells.
 - a. Make up an antibody master mix with APC-conjugated anti-IFN γ antibody diluted into FACS buffer (do not dilute into Perm2 buffer).
 - b. Aliquot 50 μ l of antibody cocktail into each well by single channel pipette. Mix well by pipetting up and down using a multichannel pipette. Allow plate to incubate on ice for 30 min to stain.
27. Wash once by adding 150 μ l FACS buffer to each well using a multichannel pipette. Centrifuge plate at 915 x *g* in a 4 °C centrifuge for 5 min.
28. Flick supernatant into a container to be discarded and blot plate on paper towels.
29. To fix, add 50 μ l per well of 2% paraformaldehyde diluted with PBS using a multichannel pipette. Cells should be fixed a minimum of 10 min on ice, but samples can be stored at 4 °C for up to 2 days without significant changes in fluorescent staining. The fixative does not need to be washed out prior to analysis.
30. Run cells on BD LSR-II.
31. Data are analyzed using FlowJo Single Cell Analysis software. Gates for IFN γ and CD107a expression are set using FMOs on a control sample and applied to all other samples collected. An example of the gating strategy and results of IFN γ and CD107a expression following media only (*i.e.* no stimulation), K562 target cell stimulation and IL-12 + IL-18 stimulation are shown (Figure 1).

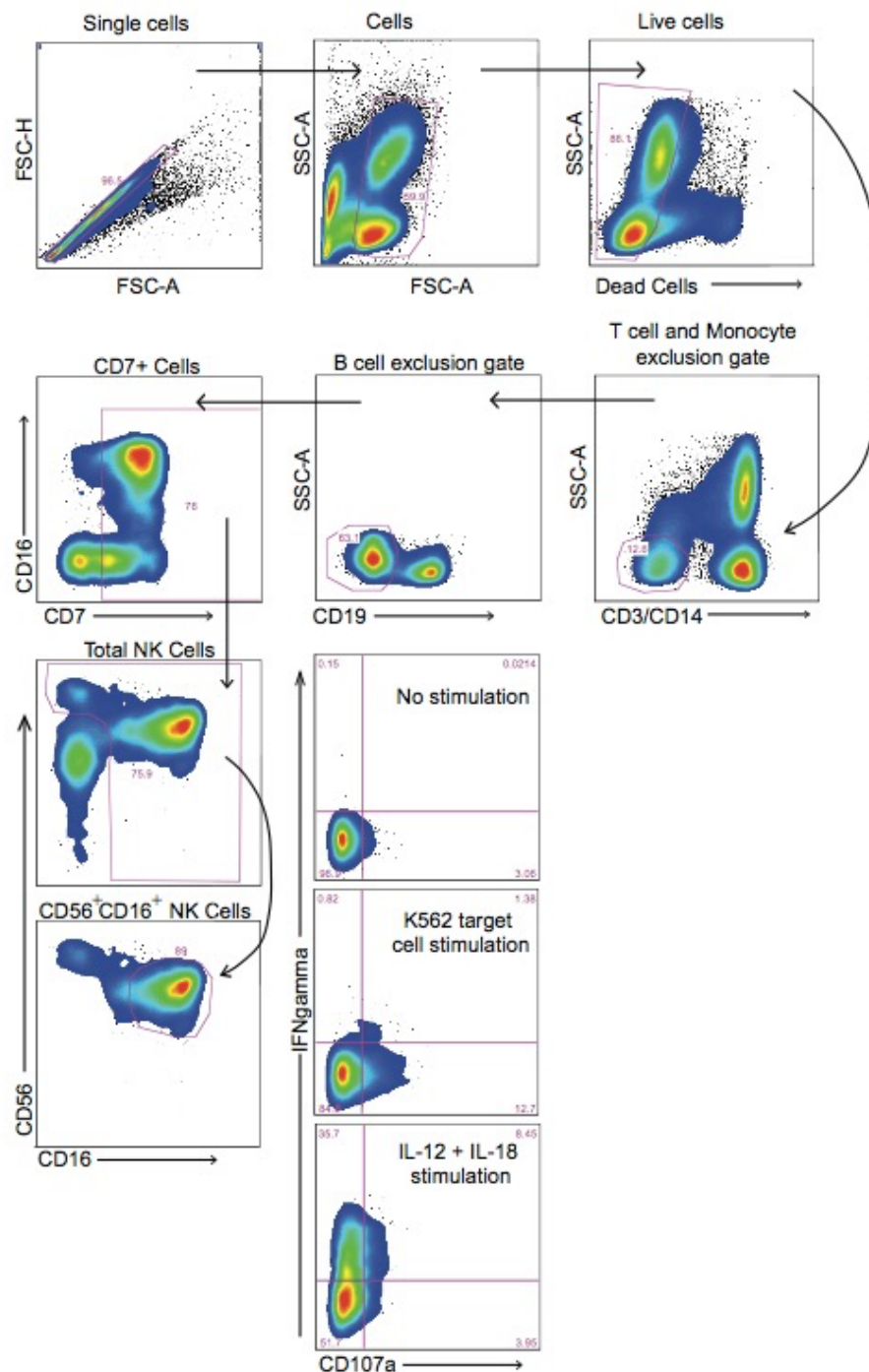


Figure 1. Representative gating strategy to identify NK cells and responses to stimulation. NK cells are identified as live lymphocytes that are CD3, CD14 and CD19 negative, but express CD7, CD56 and CD16. K562 target cell stimulation induces expression of both CD107a and IFN γ . IL-12 + IL-18 cytokine stimulation induces a robust IFN γ response from NK cells, however these cytokines do not result in degranulation as measured by CD107a expression.

Recipes

1. Complete media (make fresh each time)
 - RPMI
 - 10% Fetal Bovine Serum
 - 2 mM L-glutamine
 - Penicillin (100 Units/ml)/Streptomycin (100 μ g/ml)
 - DNase 0.01 mg/ml
2. FACS buffer (shelf life 1 month)
 - Phosphate buffer saline
 - 0.5% Bovine serum albumin
 - 2 mM Ethylenediaminetetraacetic Acid
3. Paraformaldehyde (PFA) recipe
 - Dilution of PFA made fresh each time
 - Dilute 16% stock PFA in PBS to a final concentration of 2%

Acknowledgments

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References

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