

## Intestinal Differentiation of Human ESCs

Soichiro Ogaki and Shoen Kume\*

Department of Stem Cell Biology, Kumamoto University, Kumamoto, Japan

\*For correspondence: [skume@kumamoto-u.ac.jp](mailto:skume@kumamoto-u.ac.jp)

**[Abstract]** ES cells (ESCs) are pluripotent and offer a good tool to study early embryonic development. Intestinal cells are derived from the definitive endoderm. In contrast to adult tissue stem cells, embryonic development and differentiation from ES cells have not been as well investigated in the intestine. There are four differentiated cell types of non-proliferative epithelial cells: enterocytes, goblet cells, enteroendocrine cells, and Paneth cells. Intestinal stem cells (ISCs) and progenitor cells reside in crypts, proliferate vigorously, and function as the source of differentiated epithelial cells. Here, we describe a protocol, in which differentiated cell types of the intestine are derived from human ESCs. In this protocol, we describe a protocol to differentiate mouse ES cells into Cdx2-expressing intestinal endoderm efficiently by co-culturing with M15, a mouse mesonephric cell line, and treatment with two chemical compounds. The chemical compounds used are BIO and DAPT. BIO is a Gsk3 inhibitor, that activate Wnt signaling pathway, and DAPT is a-secretase inhibitor that inhibit Notch signaling pathway. BIO and DAPT treatment yielded all representative cell lineages, enterocytes, goblet cells, enteroendocrine cells and paneth cells, to be derived from human ESCs. The protocol for human ESCs is principally very similar with that for the mouse ESCs, with some modifications.

### Materials and Reagents

1. Human embryonic stem cells (ESCs)
2. M15 cells (ECACC, catalog number: 95102517)
3. Mouse embryonic fibroblast (MEF)
4. 0.1% gelatin
5. PBS
6. Dulbecco's Modified Eagle Medium (DMEM) high glucose (Gibco®, catalog number: 11995-075)
7. DMEM low glucose (Gibco®, catalog number: 11885-084)
8. KnockOut DMEM/F12 (Gibco®, catalog number: 12660-012)
9. RPMI1640 (Gibco®, catalog number: 11875-093)
10. Fetal Bovine Serum (FBS) (Hyclone, catalog number: SH30310.03)
11. 200 mM L-glutamine (L-Gln) (Nacalai Tesque, catalog number: 16948-04)

12. 5,000 units/ml mixture of penicillin and streptomycin (PS) (Nacalai Tesque, catalog number: 26252-94)
13. 10 mM MEM Non-Essential Amino Acids Solution (NEAA) (Gibco®, catalog number: 11140-050)
14. 2-mercaptoethanol (2-ME) (Sigma-Aldrich, catalog number: M7522)
15. D-(+)-Glucose (Sigma-Aldrich, catalog number: G5146)
16. KnockOut Serum Replacement (KSR) (Gibco®, catalog number: 10828)
17. B27 supplements (Gibco®, catalog number: 17504044)
18. Recombinant human Activin A (R&D Systems, catalog number: 338-AC)
19. Recombinant human bFGF (Pepro Tech, catalog number: 100-18B)
20. BIO (Calbiochem®, catalog number: 361550)
21. DAPT (Wako Pure Chemical Industries, catalog number: 041-30983)
22. Y-27432 (Wako Pure Chemical Industries, catalog number: 688000)
23. Mitomycin C (MMC) (Sigma-Aldrich, catalog number: M4287)
24. 0.05% Trypsin-EDTA (Life Technologies, catalog number: 2014-11)
25. 0.25% Trypsin-EDTA (Life Technologies, catalog number: 25200-072)
26. EF medium (see Recipes)
27. hESC Medium (see Recipes)
28. EB medium (see Recipes)
29. hESCs Endoderm Medium (see Recipes)
30. Intestinal Medium (see Recipes)

## **Equipment**

1. 24 well plate (Corning, catalog number: 3526)
2. 10 mm dish (Corning, catalog number: 430167)
3. 60 mm dish (BD Biosciences, Falcon®, catalog number: 353004)
4. Centrifuge
5. 37 °C 5% CO<sub>2</sub> Cell culture incubator

## **Procedure**

### **A. Preparation of M15 cells**

1. 5x10<sup>6</sup> M15 cells are plated to a 100 mm dish with 10 ml EF medium.
2. M15 cells are incubated in the 5% CO<sub>2</sub> incubator at 37 °C.
3. When M15 cells are confluent (for 2-3 days), passage from a 100 mm dish to five 100 mm dishes.

- a. M15 cells are washed by PBS.
- b. M15 cells are trypsinized with 1 ml 0.05% Trypsin-EDTA at 37 °C for 5 min.
- c. M15 cells are pelleted at 190 x *g* for 5 min.
- d. M15 cells are plated to five 100 mm dishes.
4. When M15 cells are confluent, M15 cells are incubated with 10 µg/ml MMC containing EF medium in the 5% CO<sub>2</sub> incubator at 37 °C for 2 h to stop M15 cell from proliferation.
5. MMC-treated M15 cells are washed by PBS.
6. MMC-treated M15 cells are trypsinized with 1 ml 0.05% Trypsin-EDTA at 37 °C for 5 min.
7. M15 cells are pelleted at 190 x *g* for 5 min.
8. Resuspend and freeze MMC-treated M15 cells into cryovials with 22% FBS and 20% DMSO containing DMEM at 1 x 10<sup>7</sup> cells per tube at -150 °C.

#### B. Preparation of MEF

1. MEF (5x10<sup>6</sup>) are plated to a 100 mm dish with 10 ml EF medium.
2. MEF are incubated in the 5% CO<sub>2</sub> incubator at 37 °C.
3. When MEF are confluent (for 2-3 days), passage from a 100 mm dish to five 100 mm dishes.
  - a. MEF are washed by PBS.
  - b. MEF are trypsinized with 1 ml 0.05% Trypsin-EDTA at 37 °C for 5 min.
  - c. MEF are pelleted at 190 x *g* for 5 min.
  - d. MEF cells are plated to five 100 mm dishes.
4. When MEF cells are confluent, MEF are incubated with 10 µg/ml MMC containing EF medium in the 5% CO<sub>2</sub> incubator at 37 °C for 2 h. MEF are treated with MMC to stop M15 cell from proliferation.
5. MMC-treated MEF are washed by PBS.
6. MMC-treated MEF are trypsinized with 1 ml 0.05% Trypsin-EDTA at 37 °C for 5 min.
7. M15 cells are pelleted at 190 x *g* for 5 min.
8. Resuspend and freeze MMC-treated M15 cells into cryovials in 22% FBS and 20% DMSO containing DMEM at 2 x 10<sup>6</sup> cells per tube at -150 °C.

#### C. Maintenance of human ESCs

1. 1 x 10<sup>5</sup> MMC-treated MEF cells are plated to a gelatin-coated 60 mm dish with EF medium

*Note: Lower number of MEF cells is used compared to that of the mouse ESCs.*

2. MMC-treated MEF cells are incubated in the 5% CO<sub>2</sub> incubator at 37 °C overnight.
3. 5x10<sup>6</sup> Human ESCs are plated on MEF-coated dish with hESC Medium in the 5% CO<sub>2</sub> incubator at 37 °C (day 0), until passage (4-7 days).

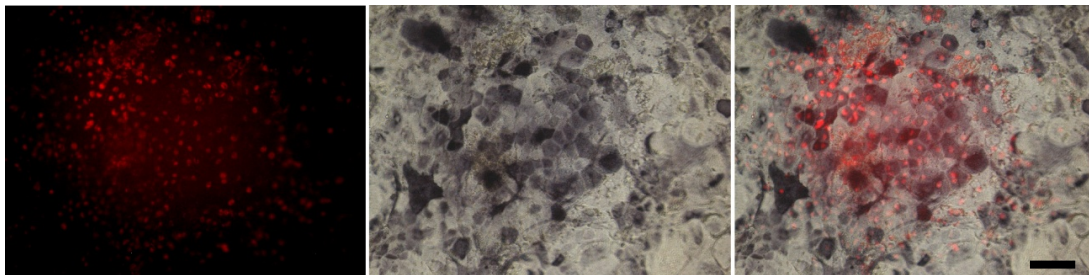
4. Medium is changed daily.

#### D. Intestinal differentiation of human ESCs

1.  $1 \times 10^7$  MMC-treated M15 cells are plated to two gelatin-coated 24 well plates in the 5% CO<sub>2</sub> incubator at 37 °C overnight.
2. ESCs plated on MEF-coated dishes (step C3) are treated hESC Medium with 10  $\mu$ M Y-27632, which is a ROCK inhibitor and is added to inhibit apoptosis, in the 5% CO<sub>2</sub> incubator at 37 °C overnight (day 1).
3. ESCs are washed by PBS.
4. ESCs are trypsinized with 1 ml 0.25% Trypsin-EDTA at 37 °C for 5 min.
5. ESC cells are pelleted at 190 x *g* for 5 min.
6. The supernatant is removed and  $1 \times 10^6$  cells/ml resuspended in EB medium.
7.  $5 \times 10^4$  ESCs in 500  $\mu$ l EB medium are plated to a M15-pre-coated well (day 0).
8. Medium is changed to hESCs Endoderm Medium at day 1, 3, 5, 7 and 9.

*Note: Human ES cells-derived definitive endodermal induction is different with that of the mouse ESCs.*

9. Medium is changed to 2 ml Intestinal Medium at day 10.
10. Medium is changed with 2 ml Intestinal Medium at every two days (Figure 1).



**Figure 1. Human ESC-derived enterocyte at day 30** Cells expressing the intestinal transcription factor CDX2 are stained positive for alkaline phosphatase (ALP), a marker for the absorptive enterocytes. Red: CDX2, Black: ALP. Scale bar indicates 100  $\mu$ m.

#### Recipes

1. EF medium  
DMEM high glucose containing 10% FBS  
2 mM L-Gln  
50 units/ml PS
2. hESC Medium  
DMEM/F12 containing 20% FBS

- 2 mM L-Gln
- 50 units/ml PS
- 100  $\mu$ M NEAA
- 100  $\mu$ M 2-ME
- 5 ng/ml bFGF
- 3. EB medium
  - DMEM high glucose containing 10% FBS
  - 2 mM L-Gln
  - 50 units/ml PS
  - 100  $\mu$ M NEAA
  - 100  $\mu$ M 2-ME
- 4. hESCs Endoderm Medium
  - RPMI1260 with 100 ng/ml Activin A
  - 2% B27 supplements
  - 2 mM L-Gln
  - 50 units/ml PS
  - 100  $\mu$ M NEAA
  - 100  $\mu$ M 2-ME
- 5. Intestinal Medium
  - DMEM low glucose with 1 mg/ml D-(+)-Glucose
  - 10% KSR
  - 2 mM L-Gln
  - 50 units/ml PS
  - 100  $\mu$ M NEAA
  - 100  $\mu$ M 2-ME
  - 5  $\mu$ M BIO
  - 10  $\mu$ M DAPT

### **Acknowledgments**

This protocol is adapted from Ogaki *et al.* (2013). This work was supported by a Grant-in-Aid (#21390280 to S.K. and #21790671 to N.S.), and in part by a Global COE grant (Cell Fate Regulation Research and Education Unit, to S.K.), and a grant from the Project for Realization of Regenerative Medicine from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) Japan.

## **References**

1. Ogaki, S., Shiraki, N., Kume, K. and Kume, S. (2013). [Wnt and Notch signals guide embryonic stem cell differentiation into the intestinal lineages.](#) *Stem Cells* 31(6): 1086-1096.