

Virulence Studies of *Clostridium difficile*

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[Abstract] *Clostridium difficile* (a Gram-positive, spore-forming, strict anaerobe) can colonize antibiotic-treated hosts (McFarland, 2008). Antibiotics alter the composition of the normal, benign microbial flora which leads to loss of colonization resistance (Wilson and Perini, 1988; Antonopoulos *et al.*, 2009). *C. difficile* spores germinate to actively growing bacteria which secrete toxins that damage the colonic epithelium (Voth and Ballard, 2005). The use of animal models of *C. difficile* disease have allowed the identification of mechanisms of colonization and virulence factors (Lyras *et al.*, 2009; Kuehne *et al.*, 2010; Francis *et al.*, 2013; Aubry *et al.*, 2012; Carter *et al.*, 2011). This protocol describes virulence studies of *C. difficile* in the hamster model of *C. difficile* infection (Bartlett *et al.*, 1978; Sambol *et al.*, 2001).

Materials and Reagents

1. Institutional Animal Care and Use Committee (IACUC)-approved animal use protocol
2. *C. difficile* spores
3. Female Syrian golden hamsters (80 g–120 g)
4. Clindamycin Injection, USP (150 mg/ml) (Hospira, catalog number: 0409-4052)
5. Dulbecco's Modified Eagle Medium (DMEM)

Equipment

1. Animal feeding needles (gavage needles)
2. 1 ml syringe
3. Dedicated BSL2 animal facility
4. Scale

Procedure

1. Weigh each hamster.
2. Gavage 100 µl clindamycin to each hamster (30 mg clindamycin / kg body weight).

Note: Dilute the stock clindamycin to the appropriate concentration using DMEM.

3. After 5 days, gavage 100 µl *C. difficile* spores.

Notes:

- a. Depending on the strain of *C. difficile* used, the infectious dose can vary. However, in several strains, inoculation with approximately 100 spores will result in lethal disease.
 - b. There are several methods to ensure reproducible infectious doses [e.g. suspending spores in phosphate buffered saline (PBS)]. Our laboratory uses Teflon-coated tubes to reduce spore adherence. Other laboratories can get reproducible infectious doses in water.
4. Closely monitor animals for signs of infection including: wet tail, poor fur coat, lethargy, loss of 15% body weight.
Note: Signs of disease are commonly observed in 2 – 4 days post infection. Be sure to closely monitor the animals for the above symptoms. Moribund animals can succumb to disease quickly.
 5. Humanly euthanize moribund animals, in accordance with your protocol that was approved by your Institutional Care and Animal Use Committee (IACUC). Score the time required to reach a moribund state.

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